



Lee Johnson v. Monsanto



3 Core Questions

1. Can Roundup be a substantial contributing factor in causing cancer?
2. Was Roundup a substantial contributing factor in causing Mr. Johnson's cancer?
3. Did Monsanto act with conscious disregard of human health?

1. Can Roundup be a substantial contributing factor in causing cancer?

Preponderance of Evidence: 50.01%



"I'm not sure, but I think so."

1. Can Roundup be a substantial contributing factor in causing cancer?

Jury Instruction

A substantial factor in causing harm is a factor that a reasonable person would consider to have contributed to the harm. It must be more than a remote or trivial factor. It does not have to be the only cause of the harm. Conduct is not a substantial factor in causing harm if the same harm would have occurred without that conduct.

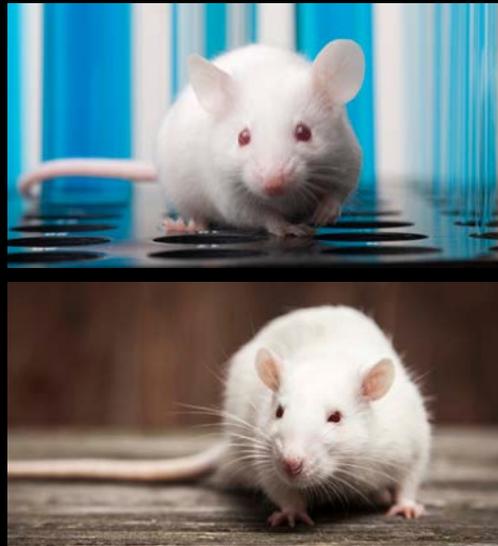
1. Can Roundup be a substantial contributing factor in causing cancer?

Three Pillars of Cancer Science

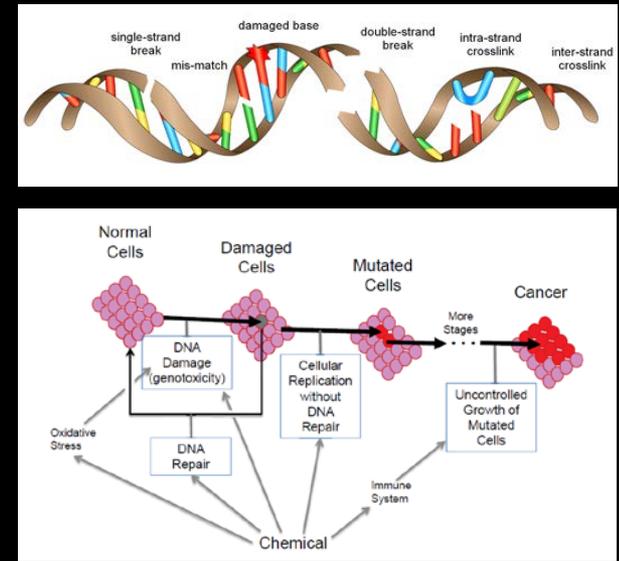
Epidemiology



Toxicology



Mechanism



Three Pillars of Cancer Science

Plaintiff's Experts Each Examined **All** the Data



Monsanto's Experts **Did Not**



(Epi only)



(Rodent only)



(AHS only)

Three Pillars of Cancer Science



Jury Instruction

The following exhibits may be admitted for the limited purpose of evaluating Monsanto's state of mind regarding the state of the science and **for no other purpose**:

1. EPA, Office of Prevention, Pesticides and Toxic Substances, Reregistration Eligibility Decision (RED) Glyphosate (Sept. 1993) [Exhibit DX2489]; and
2. EPA, Office of Pesticide Programs, Glyphosate Issue Paper: Evaluation of Carcinogenic Potential dated September 12, 2016 [Exhibit DX2481].

Three Pillars of Cancer Science



- Scientific Advisory Panel
- EPA ignored guidelines
- “Dog in the fight”
- Made mistakes before
- Glyphosate only
- Jess Rowland

Three Pillars of Cancer Science

EPA & Jess Rowland

Message

From: HEYDENS, WILLIAM F [AG/1000] [/o=Monsanto/ou=NA-1000-01/cn=Recipients/cn=230737]
on behalf of: HEYDENS, WILLIAM F [AG/1000]
Sent: 4/28/2015 2:52:30 PM
To: JENKINS, DANIEL J [AG/1920] [/o=Monsanto/ou=NA-1000-01/cn=Recipients/cn=813004]
CC: LISTELLO, JENNIFER J [AG/1000] [/o=Monsanto/ou=NA-1000-01/cn=Recipients/cn=533682]
Subject: RE: Glyphosate IARC Question

Dan,

Wow! - that's very encouraging. Thanks for the news update.

Regarding the sarcomas Jess mentions in Cheminova's mouse study, I'm assuming he is talking about the Haemangiosarcomas in high dose males (1000 mg/kg/day, the limit dose) and low numbers (1-3) of histiocytic sarcomas 'scattered' across all dose groups. These were discussed in the 2004 WHO/IARC JMAPR documents which states: "Owing to the lack of a dose-response relationship, the lack of statistical significance and the fact that the incidences recorded in this study fell within the historical ranges for control, these changes are not considered to be caused by administration of glyphosate."

Bill

From: JENKINS, DANIEL J [AG/1920]
Sent: Tuesday, April 28, 2015 9:33 AM
To: HEYDENS, WILLIAM F [AG/1000]
Cc: LISTELLO, JENNIFER J [AG/1000]
Subject: RE: Glyphosate IARC Question

Hey - cc'ing Jen

So...Jess called me out of the blue this morning:



"We have enough to sustain our conclusions. Don't need gene tox or epi. The only thing is the cheminova study with the sarcoma in mice- we have that study now and its conclusions are irrelevant (bc at limit dose...?). I am the chair of the CARC and my folks are running this process for glyphosate in reg review. I have called a CARC meeting in June..."

Also, Jess called to ask for a contact name at ATSDR. I passed on Jesslyn's email. He told me no coordination is going on and he wanted to establish some saying "if I can kill this I should get a medal". However, don't get your hopes up, I doubt EPA and Jess can kill this; but it's good to know they are going to actually make the effort now to coordinate due to our pressing and their shared concern that ATSDR is consistent in its conclusions w EPA.

Dan Jenkins
U.S. Agency Lead

Regulatory Affairs
Monsanto Company
1300 I St, NW
Suite 450 East
Washington, DC 20005




From: HEYDENS, WILLIAM F [AG/1000]
Sent: Monday, April 27, 2015 1:20 PM
To: JENKINS, DANIEL J [AG/1920]
Subject: RE: Glyphosate IARC Question

That would be great, Dan.

MONGLY00987756

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Three Pillars of Cancer Science

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MONGLY00987756

Three Pillars of Cancer Science

Epidemiology



IARC Classification

1. Sufficient
2. Limited
3. Inadequate
4. Lack of Carcinogenicity

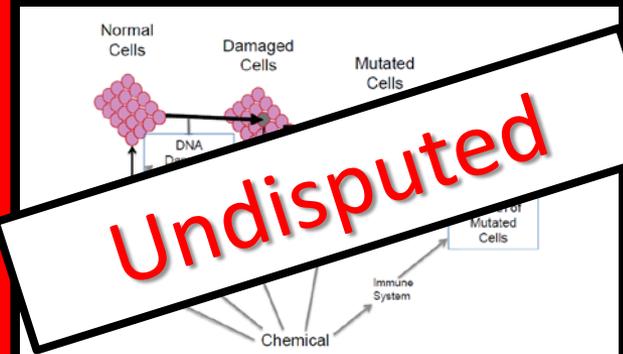
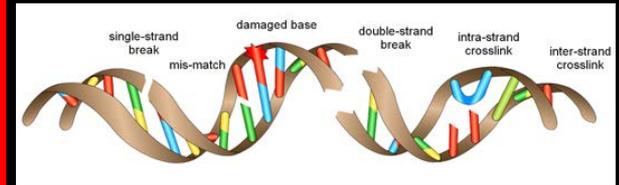
Toxicology



IARC Classification

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Mechanism



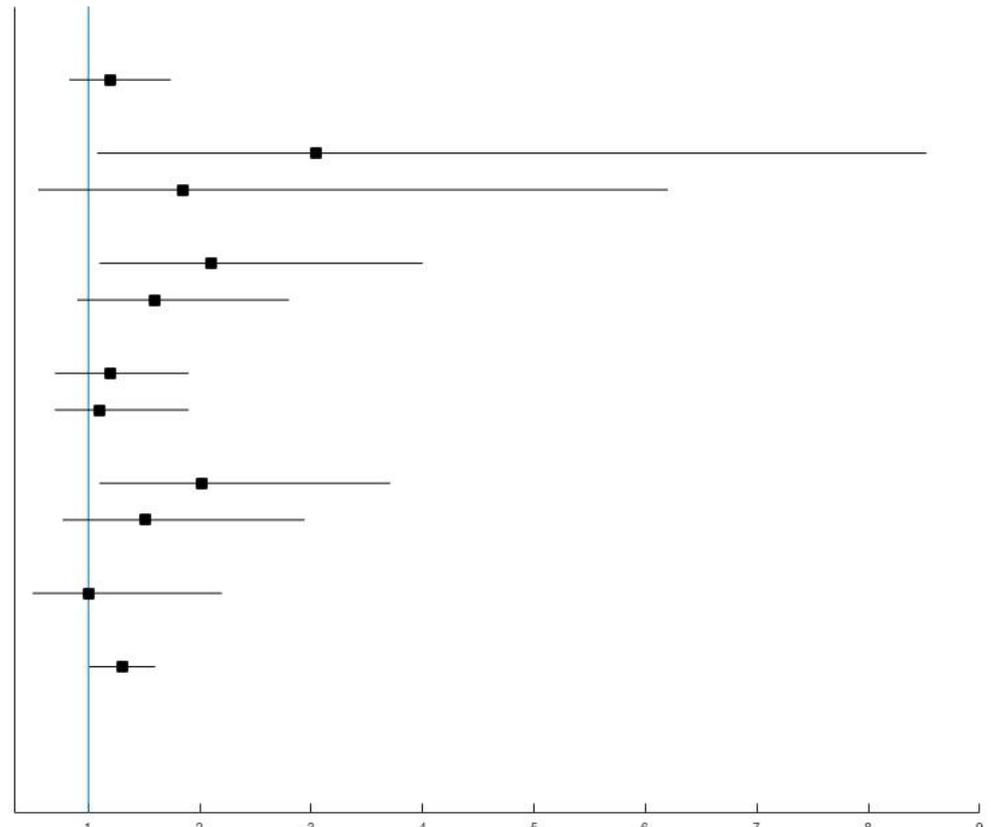
IARC Classification

1. Strong
2. Moderate
3. Weak

Three Pillars of Cancer Science

Epidemiology – Ever/Never

Study	RR	Lower	Upper
McDuffie et al. (2001) no pesticide adjustment	1.20	0.83	1.74
Hardell et al. (2002) no pesticide adjustment	3.04	1.08	8.52
Hardell et al. (2002) adjusted for pesticides	1.85	0.55	6.20
De Roos et al. (2003) adjusted for pesticides	2.10	1.10	4.00
De Roos et al. (2003) Bayesian modeling	1.60	0.90	2.80
De Roos et al. (2005) no pesticide adjustment	1.20	0.70	1.90
De Roos et al. (2005) adjusted for pesticides	1.10	0.70	1.90
Eriksson et al., (2008) no pesticide adjustment	2.02	1.10	3.71
Eriksson et al., (2008) adjusted for pesticides	1.51	0.77	2.94
Orsi et al. (2009) no pesticide adjustment	1.00	0.50	2.20
Meta-Analysis: Model 1 most adjusted analysis	1.30	1.01	1.60
Andreotti et al. (2018) not provided			



Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

- Confounding
- Proxy “bias”
- North American Pooled Project
- Agricultural Health Study

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

Confounding Is Not A Major Concern



Dr. Mucci

Q. Doctor, you said that one of the great accomplishments of epidemiology was that it helped expose that tobacco was associated with lung cancer; right?

A. Yes.

Q. And isn't it true that when that fight was happening in the epidemiology world, the tobacco companies kept saying, it's confounders?

A. Maybe. I'm sure they did, yes.

Dr. Mucci Cross, pg. 4372:3-10

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

Confounding Is Not A Major Concern



Dr. Goldstein

Message

From: GOLDSTEIN, DANIEL A [AG/1000] [/O=MONSANTO/OU=NA-1000-01/CN=RECIPIENTS/CN=527246]
Sent: 2/26/2015 8:08:31 PM
To: VICINI, JOHN L [AG/1000] [/O=MONSANTO/OU=NA-1000-01/cn=Recipients/cn=56908]; REYNOLDS, TRACEY L [AG/1000] [/O=MONSANTO/OU=Na-1000-01/cn=recipients/cn=133378]
CC: SACHS, ERIC S [AG/1000] [/O=MONSANTO/OU=NA-1000-01/cn=Recipients/cn=171736]
Subject: ACSH

While I would love to have more friends and more choices, we don't have a lot of supporters and can't afford to lose the few we have....

I am well aware of the challenges with ACSH and know Eric has valid concerns- so I can assure you I am not all starry-eyed about ACSH- they have PLENTY of warts- but:

You WILL NOT GET A BETTER VALUE FOR YOUR DOLLAR than ACSH:

Plaintiff Exhibit
0321

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Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

Confounding Is Not A Major Concern



Dr. Benbrook

Q. The ACSH, what position did it take with regards to tobacco?

A. They were one of the scientific organizations that held out to the end and argued that the science really wasn't clear about tobacco causing cancer.

Q. Talked about how too many confounding factors; right?

A. That's certainly one of the arguments that's brought up.

Dr. Benbrook Direct, pg. 3902:19-3903:10

Three Pillars of Cancer Science

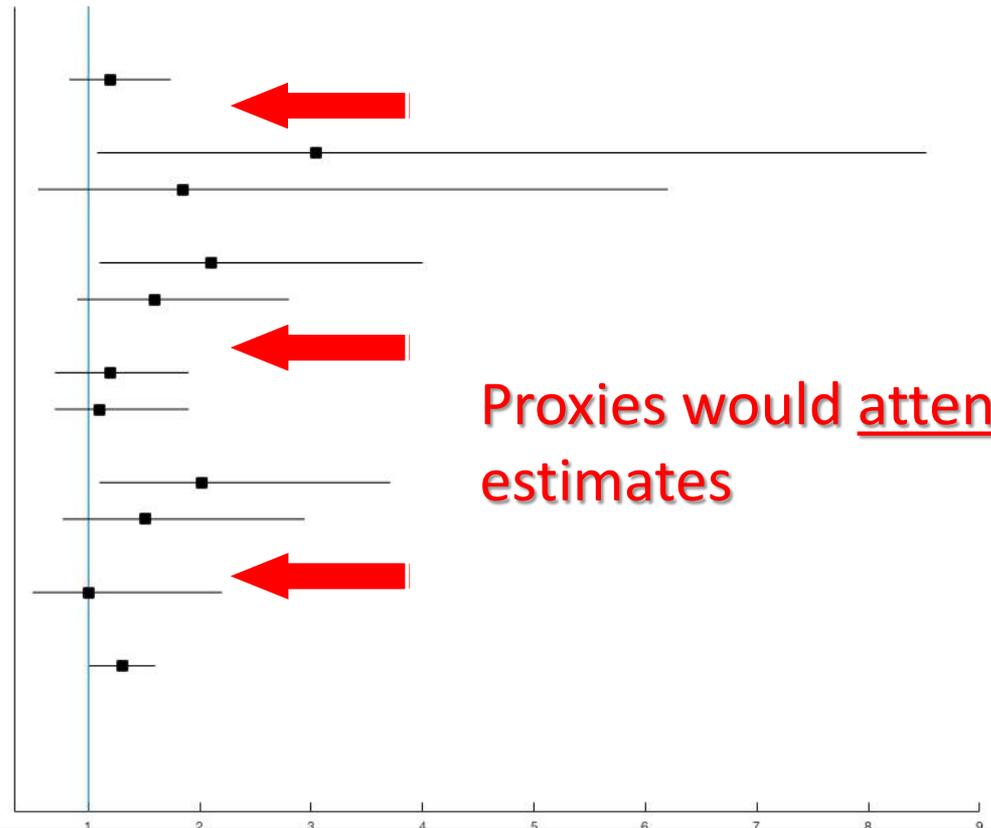
Epidemiology – Monsanto's Arguments

- **Confounding**
- Proxy “bias”
- North American Pooled Project
- Agricultural Health Study

Three Pillars of Cancer Science

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Proxies would attenuate estimates

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

- **Confounding**
- **Proxy “bias”**
- North American Pooled Project (NAPP)
- Agricultural Health Study

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

NAPP Supports IARC and NHL Association

Date of last revision: September 21, 2015

TITLE

An evaluation of glyphosate use and the risk of non-Hodgkin lymphoma major histological sub-types in the North American Pooled Project (NAPP)

AUTHORS AND AFFILIATIONS

Manisha Pahwa¹, Laura Beane Freeman², John J. Spinelli^{3,4}, Paul A. Demers^{5,6}, Aaron Blair⁷, Punam Pahwa⁸, James A. Dosman⁹, John R. McLaughlin¹⁰, Sheila Hoar Zahn¹, Kenneth P. Cantor¹, Dennis D. Weisenburger⁷, Shelley A. Harris^{1,10}

1. Occupational Cancer Research Centre, Cancer Care Ontario, Toronto, Canada
2. Division of Cancer Epidemiology and Genetics, U.S. National Cancer Institute, Bethesda, U.S.
3. British Columbia Cancer Agency Research Centre, Vancouver, Canada
4. School of Population and Public Health, University of British Columbia, Vancouver, Canada
5. Dalla Lana School of Public Health, University of Toronto, Toronto, Canada
6. Canadian Centre for Health and Safety in Agriculture, University of Saskatchewan, Saskatoon, Canada
7. Community Health and Epidemiology, College of Medicine, University of Saskatchewan, Saskatoon, Canada
8. Public Health Ontario, Toronto, Canada
9. City of Hope, Duarte, U.S.
10. Prevention and Cancer Control, Cancer Care Ontario, Toronto, Canada

TARGET JOURNAL

Occupational and Environmental Medicine

ARTICLE TYPE

Original article

KEY TERMS

Glyphosate, Lymphoma, Non-Hodgkin, Pesticides, Case-Control Studies

CORRESPONDING AUTHOR

Manisha Pahwa, Occupational Cancer Research Centre, Cancer Care Ontario, 620 University Avenue, Toronto, Ontario, Canada, M5G 2L7, Phone: 416-971-9800 ext. 3321, Fax: 416-971-6888, E-mail: manisha.pahwa@occupationalcancer.ca

WORD COUNT

Abstract: limit 250 (count 249)
Text: limit 4500 (count 622)

Page 1 of 19



Date of last revision: September 21, 2015

ABSTRACT (249)

Objectives: Glyphosate is the most frequently used herbicide worldwide. Some epidemiological studies have found positive associations between glyphosate exposure and non-Hodgkin lymphoma (NHL). This study aimed to evaluate NHL risk overall and by major histological sub-type using detailed glyphosate use metrics.

Methods: The NAPP, composed of pooled case-control studies from the U.S. and Canada, includes NHL cases (N=1690) and controls (N=5133) who provided information on pesticide use. Cases (follicular lymphoma [FL], diffuse large B-cell lymphoma [DLBCL], small lymphocytic lymphoma [SLL], other) from cancer registries and hospitals were frequency-matched to population-based controls. Logistic regression was used to estimate odds ratios (OR) and 95% confidence intervals (CI) by ever/never, duration, frequency, and lifetime days of glyphosate use. Models were adjusted for age, sex, location, proxy respondent, family history of lymphohematopoietic cancer, and personal protective equipment.

Results: Cases who ever used glyphosate (N=133) had a significantly elevated risk of NHL overall (OR=1.43, 95% CI: 1.11, 1.83). Subjects who used glyphosate for >3.5 years had increased SLL risk (OR=1.98, 95% CI: 0.89, 4.39) and those who handled glyphosate for >2 days/year had significantly elevated odds of NHL overall (OR=2.42, 95% CI: 1.48, 3.96) and DLBCL (OR=2.83, 95% CI: 1.48, 5.41). There were suggestive increases (p-trend <0.02) in risk of NHL overall, FL, and SLL with more days/year of glyphosate use.

Conclusions: Glyphosate use may be associated with increased NHL risk. Although risk differences by histological sub-type were not consistent across glyphosate use metrics, the NAPP's large sample size yielded more precise results than possible in previous studies.

Page 3 of 19

Date of last revision: September 21, 2015

misclassified PFC use data. Risks were high and unstable in this larger group due to the small number of cases in each glyphosate use category.

DISCUSSION

The objective of this study was to evaluate potential associations between glyphosate use and NHL risk in the NAPP, a large pooled dataset with detailed information about glyphosate use reported by 1690 NHL cases and 5133 controls. Glyphosate use was associated with elevated NHL risk, a finding that was consistent with previous analyses. Odds somewhat differed by histological sub-type, although there wasn't a consistent pattern across glyphosate use metrics. The novelty of this analysis and increased precision of risk estimates compared to smaller individual studies were major strengths. Yet, the limitations of this study illustrate the need for more research that can better characterize the relationship between glyphosate exposure and the development of NHL.

This report confirms previous analyses indicating increased risks of NHL in association with glyphosate exposure. The odds of NHL for glyphosate use was 1.43 (OR: 95% CI: 1.11, 1.83), a value that was situated approximately in between the risks observed in earlier analyses of the Canadian study (OR=1.26, 95% CI: 0.87, 1.80, adjusted for age and province, N=51 exposed cases) (5) and the three pooled U.S. studies (logistic regression OR=2.1, 95% CI: 1.1, 4.0, adjusted for age, study site, and other pesticides, N=26 exposed cases) (7). Further adjusting OR for the pesticides 2,4-D, dicamba, and oxabenzflorfen resulted in an attenuated risk of NHL overall in the NAPP (OR=1.23, 95% CI: 0.84, 1.81). De Roos et al. (2003) (7) used a more conservative approach, a hierarchical regression model, for assessing NHL risk in the three U.S. pooled case-control studies and found that this reduced the odds of NHL overall (OR=1.5, 95% CI: 0.9, 2.3, adjusted for age, study site, and other pesticides), a statistically significant excess of NHL was found in association with more than 2 days per year of use (OR=2.12, 95% CI: 1.20, 3.73) (5) in the Canadian study, a finding that was in agreement with our analogous pooled risk estimate for NHL (OR=2.42, 95% CI: 1.48, 3.96).

Our results are also aligned with findings from epidemiological studies of other populations that found an elevated risk of NHL for glyphosate exposure and with a greater number of days/year of glyphosate use (8), as well as a meta-analysis of glyphosate use and NHL risk (9). From an epidemiological perspective, our results were supportive of the IARC evaluation of glyphosate as a probable group 2A carcinogen for NHL (11).

The large sample size of the NAPP was conducive to analyzing NHL risks with different metrics of glyphosate use. Evaluations of dichotomous glyphosate use showed nearly universal increases in risks of NHL overall and by sub-type. But results were more varied upon further examination by duration, frequency, and lifetime days. The odds of NHL overall and by sub-type, were higher among subjects who reportedly used glyphosate more often in a year or who had greater cumulative use in their lifetime compared to unexposed subjects. Subjects who used glyphosate reported mostly initiating its use in the year 1990. Glyphosate was used by cases and controls for an average of 5 years and handled for an average of 5 days/year. The short duration of use made it challenging to calculate risks associated with longer-term usage, although the mean frequency of handling was typical of how often farmers

Page 12 of 19

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TITLE

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Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

NAPP Supports IARC and NHL Association

Date of last revision: September 21, 2015

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Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

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Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

NAPP Supports IARC and NHL Association

Because NAPP was not published, IARC could not consider this information, which would have strengthened the epidemiology data.

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

- Confounding
- Proxy “bias”
- North American Pooled Project (NAPP)
- Agricultural Health Study (AHS)

Three Pillars of Cancer Science

Epidemiology – Monsanto's Arguments

AHS Is Incapable of Detecting an Association

July 22, 1997

To the Communications Subcommittee:

At your last meeting, I was asked to provide some background thoughts on Epidemiology and the Agricultural Health Study (AHS) that you could use to build positive messages. Please find some preliminary thoughts attached.

I have put your request for background information on the agenda for the next Epidemiology Work Group meeting (August 7th). This will give you the benefit of input from a broader sphere of scientists. The Epi Work Group will be glad to entertain other requests and looks forward to assisting you in your work on the AHS.

Regards,

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Background Thoughts for the Communications Subcommittee

Farmers' health profile

Farmers are not an occupational population in obvious need of etiologic research. Their total mortality rate is 24% less than the general population rate. Their cancer mortality rate (for all cancers combined) is 16% less than the general population rate. Recent analyses show that the only cancer clearly elevated among farmers is lip cancer - believed attributable to sun exposure.

The AHS rationale

The rationale for the AHS derives from results of a number of poor studies which found associations between farming or pesticide exposure (vaguely defined) and various diseases. The AHS is intended to advance the science in this area by creating a human living laboratory for decades of research. Thus, the time horizon for definitive research is long. In the short term, the AHS investigators will work to confirm some existing theories (e.g. 2,4-D and lymphoma). But, the viability and eventual impact of the AHS will depend on the investigators' ability to generate a new class of scientific leads, most of which will be invalid. This has the potential to be disruptive for the agricultural chemical industry as new leads potentially take on a life of their own. Perhaps the best way to position the AHS is as part of a learning process. The learning process will take years to be resolved and will need to incorporate information from other research (e.g. studies of manufacturing workers) before any conclusions can be established as valid.

A definition of epidemiology

A scientific discipline that conducts studies of people to identify factors that increase or decrease human rates of disease.

The ideal study

The limitations of the AHS can be illustrated by comparison with the hypothetical ideal study. The ideal study would have the following characteristics:

- experienced investigators
- well reasoned hypotheses defined before the study
- well defined study population
- comparable exposed and comparison groups
- accurate exposure assessment
- accurate disease classification

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- comprehensive data analysis
- no systematic bias and no confounding
- good documentation
- accurate/fair write-up

How the AHS compares to the ideal study

Investigators. The key NCI investigators are experienced in agricultural research and highly regarded in the epidemiologic community. The key NIEHS investigator (Dr. Sandler) is highly regarded by epidemiologists, but she and the entire NIEHS team are inexperienced in agricultural epidemiology.

Hypotheses. Most of the diseases to be studied in the AHS have scant reasoning to link them putatively to pesticide exposure. Thus, much of the research can be termed "exploratory." That's not unusual in epidemiology, but it is unusual on this big a scale.

The downside for industry and agriculture in this approach is that exploratory research tends to yield uncertain findings. Uncertain findings, at the least, cast doubt on the safety of products. This enrages pesticide opponents, may cause the public to dictate a market change, and typically makes the manufacturer adopt a defensive stance. It would have been preferable if the AHS had a limited scope and focused more detail on a few worthy questions.

Study population. The AHS has a well defined study population. The problem with the study population, from the researchers' perspective, is that they have limited contact with pesticides (farmers report about 12 days/year of use for all pesticides). A researcher would prefer to study people with constant or daily exposure.

Comparability. Comparability is a complicated issue. The fundamental goal in epidemiologic studies is to compare the disease rate for an exposed population to the rate they would have had without exposure. This can never be done in practice. In studies like the AHS, investigators make a questionable assumption that the comparison population has the same disease rate that the exposed population would have had they not been exposed. Because of this and because of the possibility of bias discussed later, epidemiologists usually are reluctant to reach conclusions unless there is a fairly big difference in disease rates between the exposed and unexposed groups - say 50% or more. There is a strong sentiment in the epidemiologic community to dispense with this caution. We'll see how the AHS investigators treat small differences in this study.

Exposure assessment. The exposure assessment in the AHS will be inaccurate. Exposure assessment will be based on historical usage as reported by the farmer or applicator on the study questionnaire(s). There are two problems with this approach: 1. usage does not necessarily mean exposure (work practices/equipment/environmental conditions determine exposure to a large degree); 2. recall can be faulty or biased, especially when historical usage information is collected. Attempts at verification over a 3 year period have found less than 70% agreement between purchasing records and reported usage.

Inaccurate exposure classification can produce spurious results. The conventional thinking in epidemiology is that exposure misclassification will most often obscure exposure-disease relationships. More recent thinking has begun to recognize that it can also create spurious exposure-disease associations. In a study of this size, there will be some, perhaps many, spurious exposure-disease findings due to exposure misclassification.

Accurate disease classification. The AHS will have accurate disease classification for their cancer studies. In these studies, diagnoses will be determined from population based cancer registries in both states. The registries used medical records as a basis for their diagnostic information and have quality control programs in place to insure accurate diagnoses.

The non-cancer research will have less accurate disease classification. This is especially true for the initial studies where disease information is self-reported with no medical verification. Here, disease itself is not being studied, rather reports of disease are being studied.

Data analysis. NCI and NIEHS have a group of very able statisticians. We can expect a complicated analysis for most of their studies.

One important statistical issue for the AHS is the multiple comparison problem - large studies with many statistical analyses will have a number of "statistically significant" findings by chance alone. The researchers have been very vague about how they will handle the multiple comparison problem.

We also have to keep in mind that even the most sophisticated statistical analysis can't correct for other aspects of the study that are less than optimum (e.g. exposure misclassification).

Bias. Bias (really research errors or extraneous factors that favor incorrect outcome - not pre-judicial judgment)

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Investigators. The key NCI investigators are experienced in agricultural research and highly regarded in the epidemiologic community. The key NIEHS investigator (Dr. Sandler) is highly regarded by epidemiologists, but she and the entire NIEHS team are inexperienced in agricultural epidemiology.

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The downside for industry and agriculture in this approach is that exploratory research tends to yield uncertain findings. Uncertain findings, at the least, cast doubt on the safety of products. This enrages pesticide opponents, may cause the public to dictate a market change, and typically makes the manufacturers adopt a defensive stance. It would have been preferable if the AHS had a limited scope and focused more detail on a few worthy questions.

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Comparability. Comparability is a complicated issue. The fundamental goal in epidemiologic studies is to compare the disease rate for an exposed population to the rate they would have had without exposure. This can never be done in practice. In studies like the AHS, investigators make a questionable assumption that the comparison population has the same disease rate that the exposed population would have had they not been exposed. Because of this and because of the possibility of bias discussed later, epidemiologists usually are reluctant to reach conclusions unless there is a fairly big difference in disease rates between the exposed and unexposed groups - say 50% or more. There is a strong sentiment in the epidemiologic community to dispense with this caution. We'll see how the AHS investigators treat small differences in this study.

Exposure assessment. The exposure assessment in the AHS will be inaccurate. Exposure assessment will be based on historical usage as reported by the farmer or applicator on the study questionnaire(s). There are two problems with this approach: 1. usage does not necessarily mean exposure (work practices/equipment/environmental conditions determine exposure to a large degree); 2. recall can be faulty or biased, especially when historical usage information is collected. Attempts at verification over a 3 year period have found less than 70% agreement between purchasing records and reported usage.

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Farmers are not an occupational population in obvious need of etiologic research. Their total mortality rate is 24% less than the general population rate. Their cancer mortality rate (for all cancers combined) is 16% less than the general population rate. Recent analyses show that the only cancer clearly elevated among farmers is lip cancer - believed attributable to sun exposure.

The AHS rationale

The rationale for the AHS derives from results of a number of poor studies which found associations between farming or pesticide exposure (vaguely defined) and various diseases. The AHS is intended to advance the science in this area by creating a human living laboratory for decades of research. Thus, the time horizon for definitive research is long. In the short term, the AHS investigators will work to confirm some existing theories (e.g. 2,4-D and lymphoma). But, the viability and eventual impact of the AHS will depend on the investigators' ability to generate a new class of scientific leads, most of which will be invalid. This has the potential to be disruptive for the agricultural chemical industry as new leads potentially take on a life of their own. Perhaps the best way to position the AHS is as part of a learning process. The learning process will take years to be resolved and will need to incorporate information from other research (e.g. studies of manufacturing workers) before any conclusions can be established as valid.

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Impact of Pesticide Exposure Misclassification on Estimates of Relative Risks in the Agricultural Health Study

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⁷University of Iowa, Iowa City, IA
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Abstract

Background—The Agricultural Health Study (AHS) is a prospective study of licensed pesticide applicators (largely farmers) and their spouses in Iowa and North Carolina. We evaluate the impact of occupational pesticide exposure misclassification on relative risks using data from the cohort and the AHS Pesticide Exposure Study (AHS-PES).

Methods—We assessed the impact of exposure misclassification on relative risks using the range of correlation coefficients observed between measured post-application urinary levels of 2,4-dichlorophenoxyacetic acid (2,4-D) and chlorpyrifos metabolite and exposure estimates based on an algorithm from 83 AHS pesticide applications.

Results—The correlations between urinary levels of 2,4-D and chlorpyrifos metabolite and estimated exposure intensity scores from the expert-derived algorithm were about 0.4 for 2,4-D (n=64), 0.8 for liquid chlorpyrifos (n=4), and 0.6 for granular chlorpyrifos (n=12). Correlations of urinary levels with individual exposure determinants (e.g., kilograms of active ingredient used, duration of application, or number of acres treated) were lower and ranged from -0.36 to 0.19. These findings indicate that scores from an *a priori* expert-derived algorithm developed for the AHS were more closely related to measured urinary levels than the several individual exposure determinants evaluated here. Estimates of potential bias in relative risks observed in the AHS based on the correlations from the AHS-PES and the proportion of the AHS cohort exposed to various pesticides indicate that nondifferential misclassification of exposure using the algorithm would bias some estimates toward the null, but less than the misclassification associated with individual exposure determinants.

Conclusions—Based on these correlations and the proportion of the AHS cohort exposed to various pesticides, the potential bias in relative risks from nondifferential exposure misclassification is reduced when exposure estimates are based on an expert algorithm compared to estimates based on separate individual exposure determinants often used in epidemiologic

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simultaneous collection of information on exposure determinants by questionnaire or observation and measurement of urinary levels of pesticides. Estimates of exposure intensity based on self-reported activities that occurred years in the past would probably be subject to greater error. Second, the correlations between algorithm scores and urinary levels varied by pesticide in each of the three recent methodologic studies^{9,10,11} and the range was quite large, i.e., from $r=0.12$ to 0.89 . Third the impact of misclassification on estimates of relative risks is influenced by the proportion of individuals exposed because this affects the sensitivity and specificity levels. For the range of exposure misclassification noted here, it appears that the proportion of the population exposed was less important than the accuracy of the exposure assessment. This conclusion, however, is based on relatively thin data and a more complete evaluation of this issue is needed.

Some cautions about these findings are warranted. The AHS-PES monitoring study provides information on farmer owner/operators and may not be relevant for other pesticide applicators. The number of measurements on chlorpyrifos is quite small and estimates are relatively unstable. The differences between urinary levels and individual determinants and algorithm scores we observed need further evaluation to see if they are generalizable to other situations. However, these data provide useful evidence regarding the reliability of the exposure metrics used in the AHS and for the interpretation of AHS findings.

We draw several conclusions from our methodologic work in the AHS. First, the accuracy of reporting of pesticide use by farmers is comparable to that for many other factors commonly assessed by questionnaire for epidemiologic studies.^{23–25} Second, except in situations where exposure estimation is quite accurate (i.e., correlations of 0.70 or greater with true exposure) and true relative risks are 3.0 or more, pesticide misclassification may diminish risk estimates to such an extent that no association is obvious, which indicates false-negative findings might be common. Third, it appears that an algorithm that incorporates several exposure determinants into an estimate of exposure intensity predicts urinary levels better than the individual exposure determinants considered here and would result in less attenuation of relative risk estimates. This provides some confirmation of the assumption that use of algorithms will improve exposure assessment. Finally, we note that even with the reduction in power from exposure misclassification, the AHS has identified some statistically significant links between various agricultural exposures and health outcomes.^{26–28}

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The findings and conclusions in this report are those of the author(s) and do not necessarily represent the views of the National Cancer Institute, National Institute of Environmental Health Sciences, U.S. Environmental Protection Agency, or National Institute for Occupational Safety and Health.

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We thank the participants of the AHS for their contribution to this research.

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1. Breslow NE, Day NE. The analysis of case-control studies. Vol. 1. Lyon: IARC Sci Publ No. 338; 1979. Statistical methods in cancer research.
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more complete evaluation of this issue is needed.

Some cautions about these findings are warranted. The AHS/PES monitoring study provides information on farmer owner/operators and may not be relevant for other pesticide applicators. The number of measurements on chlorpyrifos is quite small and estimates are relatively unstable. The differences between urinary levels and individual determinants and algorithm scores we observed need further evaluation to see if they are generalizable to other situations. However, these data provide useful evidence regarding the reliability of the exposure metrics used in the AHS and for the interpretation of AHS findings.

We draw several conclusions from our methodologic work in the AHS. First, the accuracy of reporting of pesticide use by farmers is comparable to that for many other factors commonly assessed by questionnaire for epidemiologic studies.^{23–28} Second, except in situations where exposure estimation is quite accurate (i.e., correlations of 0.70 or greater with true exposure) and true relative risks are 3.0 or more, pesticide misclassification may diminish risks estimates to such an extent that no association is obvious, which indicates false negative findings might be common. Third, it appears that an algorithm that incorporates several exposure determinants into an estimate of exposure intensity predicts urinary levels better than the individual exposure determinants considered here and would result in less attenuation of relative risk estimates. This provides some confirmation of the assumption that use of algorithms will improve exposure assessment. Finally, we note that even with the reduction in power from exposure misclassification, the AHS has identified some statistically significant links between various agricultural exposures and health outcomes.^{29–35}

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Impact of Pesticide Exposure Misclassification on Estimates of Relative Risks in the Agricultural Health Study

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Abstract

Background—The Agricultural Health Study (AHS) is a prospective study of licensed pesticide applicators (largely farmers) and their spouses in Iowa and North Carolina. We evaluate the impact of occupational pesticide exposure misclassification on relative risks using data from the cohort and the AHS Pesticide Exposure Study (AHS-PES).

Methods—We assessed the impact of exposure misclassification on relative risks using the range of correlation coefficients observed between measured post-application urinary levels of 2,4-dichlorophenoxyacetic acid (2,4-D) and chlorpyrifos metabolite and exposure estimates based on an algorithm from 83 AHS pesticide applications.

Results—The correlations between urinary levels of 2,4-D and chlorpyrifos metabolite and estimated exposure intensity scores from the expert-derived algorithm were about 0.4 for 2,4-D (n=64), 0.8 for liquid chlorpyrifos (n=4), and 0.6 for granular chlorpyrifos (n=12). Correlations of urinary levels with individual exposure determinants (e.g., kilograms of active ingredient used, duration of application, or number of acres treated) were lower and ranged from -0.36 to 0.19. These findings indicate that scores from an *a priori* expert-derived algorithm developed for the AHS were more closely related to measured urinary levels than the several individual exposure determinants evaluated here. Estimates of potential bias in relative risks observed in the AHS based on the correlations from the AHS-PES and the proportion of the AHS cohort exposed to various pesticides indicate that nondifferential misclassification of exposure using the algorithm would bias some estimates toward the null, but less than the misclassification associated with individual exposure determinants.

Conclusions—Based on these correlations and the proportion of the AHS cohort exposed to various pesticides, the potential bias in relative risks from nondifferential exposure misclassification is reduced when exposure estimates are based on an expert algorithm compared to estimates based on separate individual exposure determinants often used in epidemiologic



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simultaneous collection of information on exposure determinants by questionnaire or observation and measurement of urinary levels of pesticides. Estimates of exposure intensity based on self-reported activities that occurred years in the past would probably be subject to greater error. Second, the correlations between algorithm scores and urinary levels varied by pesticide in each of the three recent methodologic studies^{19–21} and the range was quite large, i.e., from $r=0.12$ to 0.89. Third the impact of misclassification on estimates of relative risks is influenced by the proportion of individuals exposed because this affects the sensitivity and specificity levels. For the range of exposure misclassification noted here, it appears that the proportion of the population exposed was less important than the accuracy of the exposure assessment. This conclusion, however, is based on relatively thin data and a more complete evaluation of this issue is needed.

Some cautions about these findings are warranted. The AHS-PES monitoring study provides information on farmer owner/operators and may not be relevant for other pesticide applicators. The number of measurements on chlorpyrifos is quite small and estimates are relatively unstable. The differences between urinary levels and individual determinants and algorithm scores we observed need further evaluation to see if they are generalizable to other situations. However, these data provide useful evidence regarding the reliability of the exposure metrics used in the AHS and for the interpretation of AHS findings.

We draw several conclusions from our methodologic work in the AHS. First, the accuracy of reporting of pesticide use by farmers is comparable to that for many other factors commonly assessed by questionnaire for epidemiologic studies.^{22–25} Second, except in situations where exposure estimation is quite accurate (i.e., correlations of 0.70 or greater with true exposure) and true relative risks are 3.0 or more, pesticide misclassification may diminish risk estimates to such an extent that no association is obvious, which indicates false-negative findings might be common. Third, it appears that an algorithm that incorporates several exposure determinants into an estimate of exposure intensity predicts urinary levels better than the individual exposure determinants considered here and would result in less attenuation of relative risk estimates. This provides some confirmation of the assumption that use of algorithms will improve exposure assessment. Finally, we note that even with the reduction in power from exposure misclassification, the AHS has identified some statistically significant links between various agricultural exposures and health outcomes.^{26–28}

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Epidemiology – Monsanto's Arguments

AHS Is Incapable of Detecting an Association

IARC carefully considered the AHS data from De Roos 2005 and concluded the AHS was negative. The recent results continue to be negative and, thus, would not have affected IARC's review

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Epidemiology – Monsanto's Arguments

- Confounding
- Proxy “bias”
- North American Pooled Project (NAPP)
- Agricultural Health Study (AHS)

Three Pillars of Cancer Science

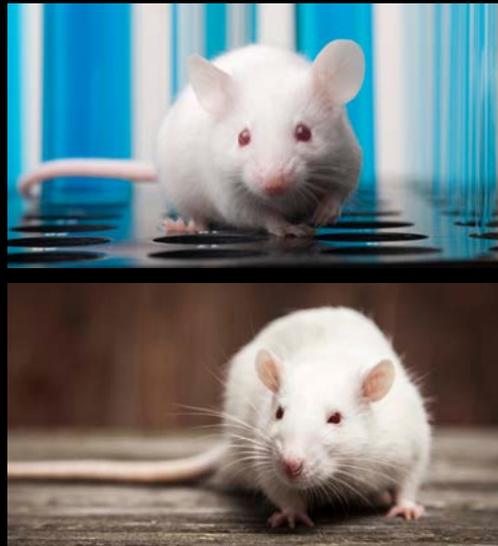
Epidemiology



IARC Classification

1. Sufficient
2. Limited
3. Inadequate
4. Lack of Carcinogenicity

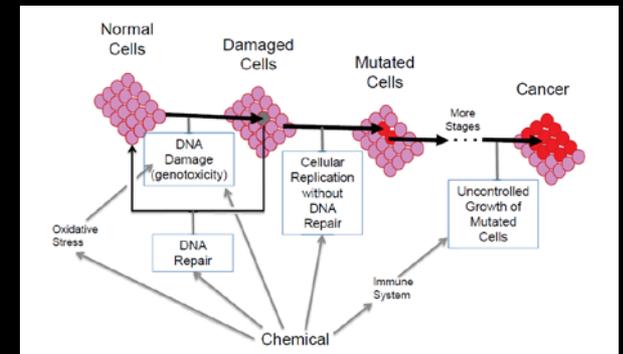
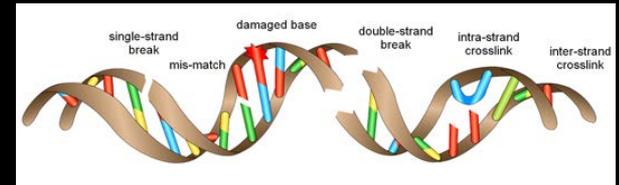
Toxicology



IARC Classification

1. Sufficient
2. Limited
3. Inadequate
4. Lack of Carcinogenicity

Mechanism



IARC Classification

1. Strong
2. Moderate
3. Weak

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Toxicology

Mice Studies - Tumor Chart

Knezevich & Hogan (1983)	Atkinson (1993) <i>Limited</i>	Sugimoto (1997)	Wood (2009)	Kumar (2001)
Kidney carcinomas or adenomas Trend Dose M / F	Malignant lymphoma Trend Dose M / F	Kidney carcinomas or adenomas Trend Dose M / F	Malignant lymphoma Trend Dose M / F	Kidney carcinomas or adenomas Trend Dose M / F
Spleen composite lymphosarcoma Trend Dose M / F	Hemangiosarcoma Trend Dose M / F	Malignant lymphoma Trend Dose M / F	Mul. malignant tumors or neoplasms Trend Dose M / F	Malignant lymphoma Trend Dose M / F
		Hemangiosarcoma Trend Dose M / F	Lung adenocarcinoma Trend Dose M / F	Hemangioma Trend Dose M / F
		Hemangioma Trend Dose M / F		
		Mul. malignant tumors or neoplasms Trend Dose M / F		
		Harderian gland adenoma Trend Dose M / F		

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Rat Studies - Tumor Chart

Lankas (1981)	Stout & Ruecker (1990)	Atkinson (1993) <i>Limited</i>	Enemoto (1997)	Suresh (1996) <i>Limited</i>	Brammer (2001)	Wood (2009)
Testicular interstitial cell tumors Trend Dose M / F	Thyroid C-Cell carcinomas or adenomas Trend Dose M / F	Thyroid follicular carcinomas or adenomas Trend Dose M / F	Kidney carcinomas or adenomas Trend Dose M / F		Hepatocellular carcinomas or adenomas Trend Dose M / F	Skin keratoacanthoma Trend Dose M / F
Thyroid C-Cell carcinomas or adenomas Trend Dose M / F	Pancreatic islet cell tumors Trend Dose M / F	Skin keratoacanthoma Trend Dose M / F	Skin keratoacanthoma Trend Dose M / F			Mammary gland carcinomas or adenomas Trend Dose M / F
Pancreatic islet cell tumors Trend Dose M / F	Hepatocellular carcinomas or adenomas Trend Dose M / F		Basal cell tumors Trend Dose M / F			Pituitary adenomas Trend Dose M / F
	Adrenal cortical carcinomas Trend Dose M / F					
	Skin keratoacanthoma Trend Dose M / F					

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Pancreatic islet cell tumors Trend Dose M / F	Hepatocellular carcinomas or adenomas Trend Dose M / F		Basal cell tumors Trend Dose M / F			Pituitary adenomas Trend Dose M / F
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Three Pillars of Cancer Science

Toxicology

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		Hemangioma Trend Dose M / F		
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Toxicology

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		Hemangiosarcoma Trend Dose M/F	Lung adenocarcinomas Trend Dose M/F	

Sufficient

Studies - Tumor Chart

Lankas (1981)	Stout & Ruecker (1990)	Atkinson (1993) <i>Limited</i>	Enemoto (1997)	Suresh (1996) <i>Limited</i>	Brammer (2001)	Wood (2009)
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	Skin keratoacanthoma Trend Dose M/F					

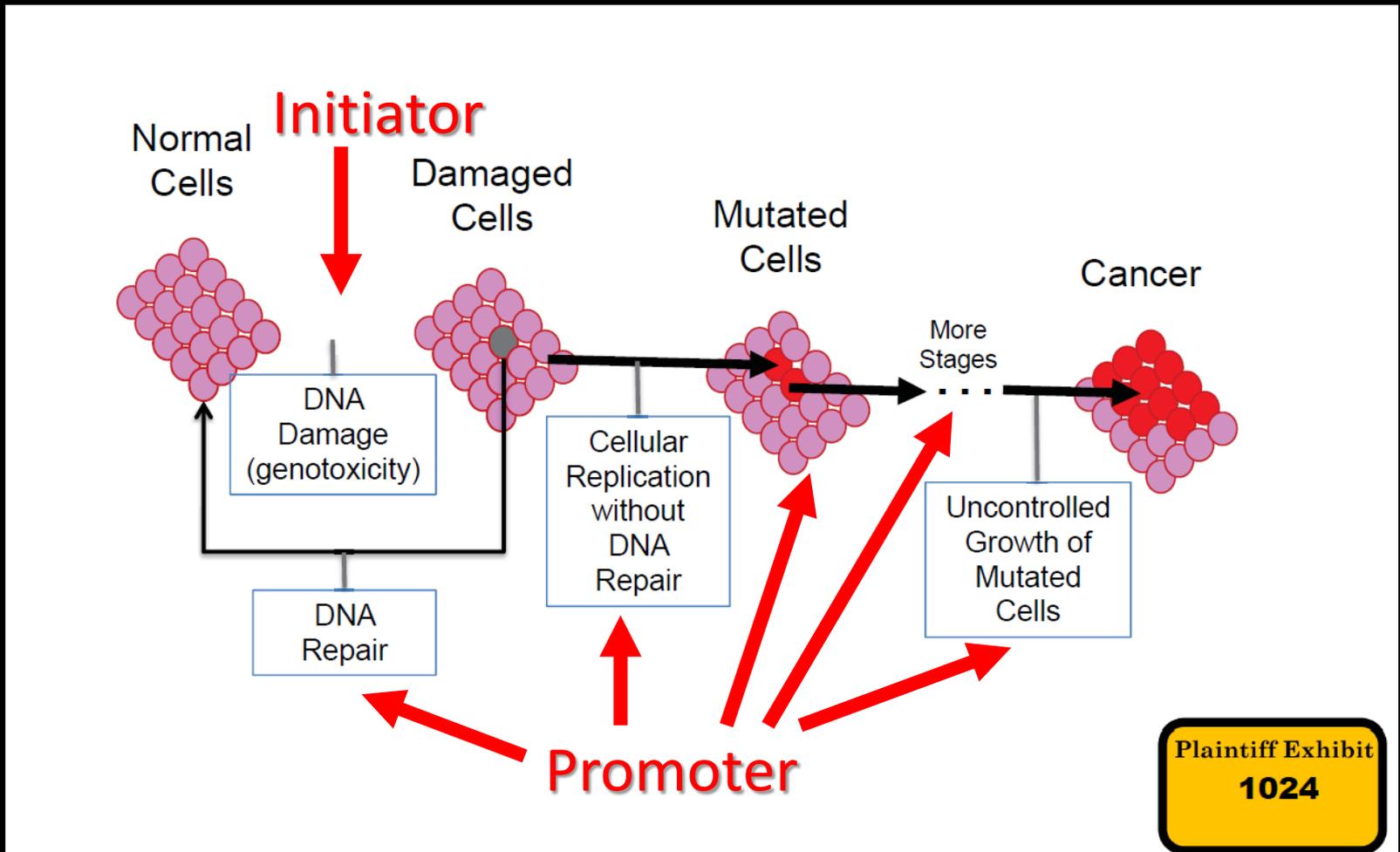


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Toxicology



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Toxicology



Studies on glyphosate-induced carcinogenicity in mouse skin: A proteomic approach

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ABSTRACT

Glyphosate is a widely used broad spectrum herbicide, reported to induce various toxic effects in non-target species, but its carcinogenic potential is still unknown. Here we showed the carcinogenic effects of glyphosate using 2-stage mouse skin carcinogenesis model and proteomic analysis. Carcinogenicity study revealed that glyphosate has same promoting activity. Proteomic analysis using 2-dimensional gel electrophoresis and mass spectrometry showed that 22 spots were differentially expressed (>2 fold) on glyphosate, 7, 12-dimethylbenzylisothiazolone (DMBA) and 12-O-tetradecanoylphorbol-13-acetate (TPA) application over untreated control. Among them, 9 proteins (translational elongation factor eIF1 α , alpha chain, catalase, ankyrin II, annexin II, cyclophilin, fibronectin, alpha-VIII antibody, pectatelectin-2, superoxide dismutase [Cu-Zn], stefin A), and calgranulin-B were common and showed similar expression pattern in glyphosate and TPA-treated mouse skin. These proteins are known to be involved in several key processes like apoptosis and growth-inhibition, anti-oxidant responses, etc. The up-regulation of cyclophilin, calgranulin-B and down-regulation of superoxide dismutase [Cu-Zn] was further confirmed by immunoblotting, indicating that these proteins can be good candidate biomarkers for skin carcinogenesis induced by glyphosate. Altogether, these results suggested that glyphosate has tumor promoting potential in skin carcinogenesis and its mechanism seems to be similar to TPA.

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1. Introduction

Herbicides, used extensively for controlling pest and destroying weeds are ubiquitous contaminants accumulating in environment and hence humans get unavoidably exposed to these pesticides. About 3 billion tons of pesticides are used every year, on agricultural crops worldwide [1]. In some cases, even short term exposure of the pesticides can make impact on human health. Apart from the other toxic effects, pesticides are reported to cause genotoxicity/carcinogenicity also. Some pesticides have been classified as carcinogens by the United States Environmental Protection Agency (USEPA) [2] and International Agency for Research on Cancer (IARC) [3]. Glyphosate, N-(phosphonomethyl) glycine, commonly sold as

a commercial formulation named, Roundup is a widely used herbicide on both cropland and non-cropland areas [4]. The potential activity of glyphosate is through competitive inhibition of the *5-enolpyruvyl-shikimate-phosphate* synthase, an enzyme essential to the synthesis of aromatic amino acids in plants [5]. Toxicological profile of glyphosate, showed that it is a comparatively safe herbicide for animals [6]. Glyphosate shows also with its formulation products, such as surfactants and permeabilizing agents is usually considered to be harmless under both normal usage and chronic exposure [4]. In 1991, USEPA categorized this compound into class I, which means that it is probably not carcinogenic to humans [7]. Despite these reports, some case-control studies suggested an association between glyphosate exposure and the risk of non-

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Hodgkin's lymphoma [8,9]. In another study, both technical grade glyphosate and Roundup were shown to cause a rapid increase in cell division in human breast cancer cells [10]. Glyphosate has also been shown as a skin irritant [11]. Regarding the genotoxic potential, glyphosate exposure to human lymphocytes *in vitro* resulted in increased sister chromatid exchanges [12], chromosomal aberrations [13], and indicators of oxidative stress [14]. A recent study from our laboratory also showed the clastogenic effects of glyphosate in bone marrow cells of Swiss albino mice [15]. These reports prompted us to investigate its carcinogenic effect in long-term animal bioassay.

To evaluate toxicity/carcinogenicity induced by physical and chemical agents including pesticides, various test systems have been employed in bacteria, rodents and mammalian cells [16–18]. Each of these environmental challenges causes changes in DNA conformation, alterations in the levels of mRNA and protein expression, and post-translational modifications such as phosphorylation and glycosylation of proteins specific to each stressor [19]. In recent years, there has been considerable interest in linking carcinogenic/oxidative responses to gene and protein expression. Toxicoproteomics has received a lot of attention as a valuable tool to search reliable early predictive toxicity markers in response to environmental stimuli [20]. Two-dimensional gel electrophoresis (2-DE) coupled with mass spectrometry (MS), a high-throughput technique allows proteins of interest to be identified by their expression and/or modification pattern rather than using the traditional approach of translating gene expression data. Biomarkers can be used to identify causal associations and to make better quantitative estimates of these associations at relevant levels of exposure [21]. Yamamoto et al. [22] have utilized proteomic approach to identify potential biomarker candidates of hepatocellular carcinoma in rat liver.

Skin is the largest organ in the body and dermal contact is one of the most probable routes of human exposure to pesticides. Thus, mouse skin model represents a logical experimental choice [23]. As the long-term bioassay for carcinogenicity is expensive, time consuming and involves a large number of animals and ethical issues, development of biomarkers after short-term exposure are needed. The present investigation was carried out to study the carcinogenic potential of glyphosate and to identify differentially expressed proteins, using 2-DE and MS analysis after treatment with glyphosate, a known tumor promoter, 12-O-tetradecanoylphorbol-13-acetate (TPA) and tumor initiator, 7, 12-dimethylbenzylisothiazolone (DMBA) in mouse skin. Altered proteins identified through proteomic approach in our study may be potentially useful as early biomarkers, to detect the adverse effects of glyphosate.

2. Materials and methods

2.1. Materials

The commercial formulation of the herbicide glyphosate (N-phosphonomethyl glycine) Roundup Original Concentrate 41%, POEA +15%—Monsanto Company, St. Louis, MO, USA was used, which contains 360 g/l glyphosate acid equivalent

as the isopropylamine salt and was procured from local market. Immobilized pH gradient (IPG) strips and 0.5% pH3–10/IC buffer were purchased from Bio-Rad Laboratories (Hercules, CA, USA). DMBA, TPA, CHAPS, DTT, and beta-actin (clone AC-20) antibody were from Sigma-Aldrich (Missouri, USA). DNase/RNase were from Bangalore Genei (Bangalore, India). The rest of the chemicals used in the study were of analytical grade of purity and procured locally.

2.2. Animals and treatments

2.2.1. Carcinogenicity study

Male Swiss albino mice (23–25 g body weight [bw]) were taken from Indian Institute of Toxicology Research (IITR) animal breeding colony and acclimatized for 1 week. The ethical approval for the experiment was obtained from institutional ethical committee. The animals were kept under standard laboratory conditions (temperature 23 ± 2 °C, relative humidity 55–58%) and were fed with synthetic pellet basal diet (Ashirwad, Chandigarh, India) and tap water ad libitum. Animals were randomly divided into 8 groups of 20 animals each. Hair were clipped in the dorsal region with proper care in an area of 2 cm² using electrical clippers, not lubricated with oil or grease. The long term treatment was given as described earlier [24]. Briefly,

- Group I: Untreated control (No treatment).
- Group II: Glyphosate alone (25 mg/kg bw, topically 3 times per week).
- Group III: DMBA + TPA (Single topical application of DMBA, 52 µg/mouse followed 1 week later by thrice a week application of TPA, 5 µg/mouse).
- Group IV: Glyphosate (25 mg/kg bw) followed 1 week later by TPA application as in group III.
- Group V: Glyphosate (25 mg/kg bw) followed 1 week later by application of glyphosate, 25 mg/kg bw for 3 weeks [total of 9 applications], followed 1 week later by TPA application as in group III.
- Group VI: DMBA (Single topical application of DMBA, 52 µg/mouse).
- Group VII: TPA (Thrice a week topical application of TPA, 5 µg/mouse).
- Group VIII: DMBA + glyphosate (Single topical application of DMBA [52 µg in group III], followed 1 week later by topical treatment of glyphosate, 25 mg/kg bw thrice per week).

Vehicle for glyphosate, DMBA and TPA were 50% ethanol and acetone respectively.

Animals from all the groups were examined every week for gross morphological changes including body weight changes, development and volume of squamous cell papilloma (tumors) locally on the skin during the entire study period, and tumors larger than 1 mm diameter, were included in the total number of tumors. Tumor volume per tumor bearing mouse was calculated in each group using formula $V = D_1 \times D_2^2 \times \pi / 6$ (where D_1 = bigger dimension and D_2 = smaller dimension). All the surviving animals were sacrificed at the end of the study period, i.e. 32 weeks for complete carcinogenic, tumor initiating and promoting studies.

Table 1—Role of carcinogenic, tumor initiating and tumor promoting effect of glyphosate in mouse skin model of carcinogenesis.

Groups	Treatment	Number of animals with tumors	1st induction of tumors (in days)	% of animals with tumors	Total number of tumors	Avg. no. of tumors/mouse (mean ± SD)	Avg. tumor volume (mm ³) (mean ± SD)
I	Untreated	0/20	—	—	—	—	—
II	Glyphosate	0/20	—	—	—	—	—
III	DMBA + TPA	20/20	53	100	154	7.6 ± 1.1	36.4 ± 5.1
IV	Glyphosate (25 mg/kg bw) + TPA	0/20	—	—	—	—	—
V	Glyphosate (25 mg/kg bw) + DMBA	0/20	—	—	—	—	—
VI	TPA	0/20	—	—	—	—	—
VIII	DMBA + glyphosate	8/20	130	60	73	2.8 ± 0.9	26.2 ± 4.8

TPA = DMBA versus untreated group (ANOVA test, 4-stage dose, n = multiple dose). Details of treatment are provided in Materials and Methods section.

number of tumors was 7.6 ± 1.1 in group III. However, in group VII, it was 2.8 ± 0.9 (Fig. 1C, Table 1). These tumors were initiated as a minute wart like growth, which progressed during the course of experiment and average tumor volume was 36.4 ± 5.1 mm³ in group III and 26.2 ± 4.8 mm³ in group VIII (Fig. 1D, Table 1). These results clearly indicate significant tumor promoting potential of glyphosate in mouse skin model of carcinogenesis.

3.2. Protein expression profile

Using 2-DE, comparisons of differentially expressed proteins were made in mouse skin following topical treatment with glyphosate (50 mg/kg bw/mouse), TPA (10 µg/mouse) and DMBA (104 µg/mouse) with untreated mouse skin individually, using PDQuest 7.0.0 2-D gel analysis software. Representative 2-DE maps are shown in Fig. 2. Image matching derived from 4 groups showed a total of ~2600 spots. Out of these, 22 spots were differentially expressed, exhibiting > 2 fold change between values of treated and control animals (Fig. 2). These spots were excised from the gels and analyzed using MALDI-TOF/MS mass spectrometer. PMF from the proteins was obtained and the resulting spectra were used to identify the proteins with the Mascot search program. Protein spots that appeared more than once, were considered as the same protein and assigned the same number. These identified proteins were categorized according to their molecular functions (Table 2), biological functions and subcellular localization (Fig. 3A and B) as referred to SWISS-PROT database. Protein spots nos. 7 and 18–1, 16–2 were up-regulated and spot no. 13 was down-regulated by glyphosate and TPA treatment (Fig. 4). Related fingerprint mass spectra of calyculin, calgranulin-B and SOD are shown in Fig. 5.

3.3. Protein expression profile in glyphosate and TPA-treated mouse skin

Substantially common and differentially expressed protein spots among glyphosate and TPA-treated skin tissues were quantitatively analyzed individually. Comparison between the gels of glyphosate and TPA revealed that 13 specific proteins spots (1, 2, 5, 6, 1, 6, 2, 7, 8, 11, 12, 13, 15, 18–1, and 18–2)

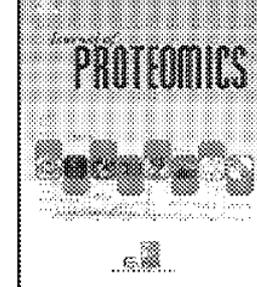
from a total of 22 spots were showing the similar expression pattern. Among the selected and identified proteins with statistically significant altered expression (p < 0.05), we focused on the proteins involved in apoptosis and growth-inhibition, anti-oxidation, energy metabolism, angiogenesis, calcium binding and protein biosynthesis processes. These proteins are translation elongation factor eIF1 α alpha chain (eIF1A), catalase, ankyrin II (CA II), annexin II, cyclophilin, fibronectin and alpha-VIC1 antibody, peroxiredoxin 2 (PRX II), superoxide dismutase [Cu-Zn] (SOD 1), stefin A, and calgranulin-B (Fig. 6, Table 3).

3.4. Protein expression profile in glyphosate and DMBA treated mouse skin

Among the 22 differentially expressed protein spots, 4 specific spots (1, 2, 7, 11, and 12) were showing the similar expression pattern between the gels of glyphosate and DMBA treated skin tissues. These proteins are eIF1A, CA II, fibronectin and alpha-VIC1 antibody and PRX II (Fig. 6, Table 3).

3.5. Cluster analysis of differentially expressed proteins in control, glyphosate, TPA and DMBA treated mouse skin

To understand the carcinogenic activity of glyphosate in mouse skin based on the level of protein expression information generated on 2-DE gels hierarchical cluster analysis was applied. The analysis facilitated the visualization of groupings based on the protein expression changes, potentially showing the relationship between glyphosate and TPA, which further leads support to their tumor promoting activity. A hierarchical clustering map is generated with the differentially expressed protein spots. The analysis showed 2 major clusters, one cluster includes TPA and glyphosate, where majority of the altered protein expression was recorded and the other cluster includes DMBA and control having comparatively low number of altered proteins (Fig. 7). Moreover, calyculin and calgranulin-B were present only in the cluster of glyphosate and TPA whereas SOD 1 is higher in DMBA and control cluster in comparison to other cluster.



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Three Pillars of Cancer Science

Toxicology



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1. Introduction

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Hodgkin's lymphoma [8,9]. In another study, both technical grade glyphosate and Roundup were shown to cause a rapid increase in cell division in human breast cancer cells [10]. Glyphosate has also been shown as a skin irritant [11]. Regarding the genotoxic potential, glyphosate exposure to human lymphocytes *in vitro* resulted in increased sister chromatid exchanges [12], chromosomal aberrations [13], and indicators of oxidative stress [14]. A recent study from our laboratory also showed the clastogenic effects of glyphosate in bone marrow cells of Swiss albino mice [15]. These reports prompted us to investigate its carcinogenic effect in long-term animal bioassay.

To evaluate toxicity/carcinogenicity induced by physical and chemical agents including pesticides, various test systems have been employed in bacteria, rodents and mammalian cells [16–18]. Each of these environmental challenges causes changes in DNA conformation, alterations in the levels of mRNA and protein expression, and post-translational modifications such as phosphorylation and glycosylation of proteins specific to each stressor [19]. In recent years, there has been considerable interest in linking carcinogenic/oxidative responses to gene and protein expression. Toxicoproteomics has received a lot of attention as a valuable tool to search reliable early predictive toxicity markers in response to environmental stimuli [20]. Two-dimensional gel electrophoresis (2-DE) coupled with mass spectrometry (MS), a high-throughput technique allows proteins of interest to be identified by their expression and/or modification pattern rather than using the traditional approach of translating gene expression data. Biomarkers can be used to identify causal associations and to make better quantitative estimates of these associations at relevant levels of exposure [21]. Yamamoto et al. [22] have utilized proteomic approach to identify potential biomarker candidates of hepatocellular carcinoma in rat liver.

Skin is the largest organ in the body and dermal contact is one of the most probable routes of human exposure to pesticides. Thus, mouse skin model represents a logical experimental choice [23]. As the long-term bioassay for carcinogenicity is expensive, time consuming and involves a large number of animals and ethical issues, development of biomarkers after short-term exposure are needed. The present investigation was carried out to study the carcinogenic potential of glyphosate and to identify differentially expressed proteins, using 2-DE and MS analysis after treatment with glyphosate, a known tumor promoter, 12-O-tetradecanoylphorbol-13-acetate (TPA) and tumor initiator, 7, 12-dimethylbenzylglutathione (DMBA) in mouse skin. Altered proteins identified through proteomic approach in our study may be potentially useful as early biomarkers, to detect the adverse effects of glyphosate.

2. Materials and methods

2.1. Materials

The commercial formulation of the herbicide glyphosate (N-phosphonomethyl glycine) Roundup Original Concentrate 41%, POEA +15%—Monsanto Company, St. Louis, MO, USA was used, which contains 360 g/l glyphosate acid equivalent

as the isopropylamine salt and was procured from local market. Immobilized pH gradient (IPG) strips and 0.5% pH3–10DQ buffer were purchased from Bio-Rad Laboratories (Berkeley, CA, USA). DMBA, TPA, CHAPS, DTT, and beta-actin (clone AC-20) antibody were from Sigma-Aldrich (Missouri, USA). DNase/RNase were from Bangalore Genei (Bangalore, India). The rest of the chemicals used in the study were of analytical grade of purity and procured locally.

2.2. Animals and treatments

2.2.1. Carcinogenicity study

Male Swiss albino mice (23–25 g body weight [bw]) were taken from Indian Institute of Toxicology Research (IITR) animal breeding colony and acclimatized for 1 week. The ethical approval for the experiment was obtained from institutional ethical committee. The animals were kept under standard laboratory conditions (temperature 23 ± 2 °C, relative humidity 55–58%) and were fed with synthetic pellet basal diet (Ashirwad, Chandigarh, India) and tap water ad libitum. Animals were randomly divided into 8 groups of 20 animals each. Hair were clipped in the dorsal region with proper care in an area of 2 cm² using electrical clippers, not lubricated with oil or grease. The long term treatment was given as described earlier [24]. Briefly,

- Group I: Untreated control (No treatment).
- Group II: Glyphosate alone (25 mg/kg bw, topically 3 times per week).
- Group III: DMBA + TPA (Single topical application of DMBA, 52 µg/mouse followed 1 week later by thrice a week application of TPA, 5 µg/mouse).
- Group IV: Glyphosate (25 mg/kg bw) followed 1 week later by TPA application as in group III.
- Group V: Glyphosate (25 mg/kg bw) followed 1 week later by TPA application as in group III.
- Group VI: DMBA (Single topical application of DMBA, 52 µg/mouse).
- Group VII: TPA (Thrice a week topical application of TPA, 5 µg/mouse).
- Group VIII: DMBA + glyphosate (Single topical application of DMBA [52 µg in group III], followed 1 week later by topical treatment of glyphosate, 25 mg/kg bw thrice per week).

Vehicle for glyphosate, DMBA and TPA were 50% ethanol and acetone respectively.

Animals from all the groups were examined every week for gross morphological changes including body weight changes, development and volume of squamous cell papilloma (tumors) locally on the skin during the entire study period, and tumors larger than 1 mm diameter, were included in the total number of tumors. Tumor volume per tumor bearing mouse was calculated in each group using formula $V = D_1^2 \times D_2^2 \times \pi$ (where D_1 = bigger dimension and D_2 = smaller dimension). All the surviving animals were sacrificed at the end of the study period, i.e. 32 weeks for complete carcinogenic, tumor initiating and promoting studies.

Table 1—Role of carcinogenic, tumor initiating and tumor promoting effect of glyphosate in mouse skin model of carcinogenesis.

Groups	Treatment	Number of animals with tumors (n days)	1st induction % of animals (n days)	% of animals with tumors	Total number of tumors	Avg. no. of tumors/mouse (mean ± SD)	Avg. tumor volume (mm ³) (mean ± SD)
I	Untreated	0/20	0	0	0	0	0
II	Glyphosate	0/20	0	0	0	0	0
III	DMBA + TPA	22/25	53	100	154	7.0 ± 1.1	36.4 ± 5.1
IV	Glyphosate (25 mg/kg bw) + TPA	0/20	0	0	0	0	0
V	Glyphosate (25 mg/kg bw) + DMBA	0/20	0	0	0	0	0
VI	DMBA	0/20	0	0	0	0	0
VII	TPA	0/20	0	0	0	0	0
VIII	DMBA + glyphosate	8/20	130	60	73	2.8 ± 0.9	26.2 ± 4.8

TPA = DMBA versus untreated group (ANOVA test, 4-stage dose, n = multiple dose). Details of treatment are provided in Materials and Methods section.

number of tumors was 7.8 ± 1.1 in group III. However, in group VII, it was 2.8 ± 0.9 (Fig. 1C, Table 1). These tumors were initiated as a minute wart like growth, which progressed during the course of experiment and average tumor volume was 36.4 ± 5.1 mm³ in group III and 26.2 ± 4.8 mm³ in group VIII (Fig. 1D, Table 1). These results clearly indicate significant tumor promoting potential of glyphosate in mouse skin model of carcinogenesis.

3.2. Protein expression profile

Using 2-DE, comparisons of differentially expressed proteins were made in mouse skin following topical treatment with glyphosate (50 mg/kg bw/mouse), TPA (10 µg/mouse) and DMBA (104 µg/mouse) with untreated mouse skin individually, using PDQuest 7.0.0 2-D gel analysis software. Representative 2-DE maps are shown in Fig. 2. Image matching derived from 4 groups showed a total of ~2600 spots. Out of these, 22 spots were differentially expressed, exhibiting >2 fold change between values of treated and control animals (Fig. 2). These spots were excised from the gels and analyzed using MALDI-TOF/MS mass spectrometer. PMF from the proteins was obtained and the resulting spectra were used to identify the proteins with the Mascot search program. Protein spots that appeared more than once, were considered as the same protein and assigned the same number. These identified proteins were categorized according to their molecular functions (Table 2), biological functions and subcellular localization (Fig. 3A and B) as referred to SWISS-PROT database. Protein spots nos. 7 and 18–1, 18–2 were up-regulated and spot no. 13 was down-regulated by glyphosate and TPA treatment (Fig. 4). Related fingerprint mass spectra of calyculin, calgranulin-B and SOD are shown in Fig. 5.

3.3. Protein expression profile in glyphosate and TPA-treated mouse skin

Substantially common and differentially expressed protein spots among glyphosate and TPA-treated skin tissues were quantitatively analyzed individually. Comparison between the gels of glyphosate and TPA revealed that 13 specific proteins spots (1, 2, 3, 6, 1, 6, 2, 7, 8, 11, 12, 13, 15, 18–1, and 18–2)

from a total of 22 spots were showing the similar expression pattern. Among the selected and identified proteins with statistically significant altered expression (p < 0.05), we focused on the proteins involved in apoptosis and growth-inhibition, anti-oxidation, energy metabolism, angiogenesis, calcium binding and protein biosynthesis processes. These proteins are translation elongation factor eIF1 α alpha chain (eIF1A), catalase, ankyrin II (CA II), annexin II, cyclophilin, fibronectin, alpha-VIC1 antibody, pectatelectin-2 (PEX II), superoxide dismutase [Cu-Zn] (SOD I), stefin A, and calgranulin-B (Fig. 6, Table 3).

3.4. Protein expression profile in glyphosate and DMBA treated mouse skin

Among the 22 differentially expressed protein spots, 4 specific spots (1, 3, 7, 1, and 12) were showing the similar expression pattern between the gels of glyphosate and DMBA treated skin tissues. These proteins are eIF1A, CA II, fibronectin, alpha-VIC1 antibody and PEX II (Fig. 6, Table 3).

3.5. Cluster analysis of differentially expressed proteins in control, glyphosate, TPA and DMBA treated mouse skin

To understand the carcinogenic activity of glyphosate in mouse skin based on the level of protein expression information generated on 2-DE gels hierarchical cluster analysis was applied. The analysis facilitated the visualization of groupings based on the protein expression changes, potentially showing the relationship between glyphosate and TPA, which further leads support to their tumor promoting activity. A hierarchical clustering map is generated with the differentially expressed protein spots. The analysis showed 2 major clusters, one cluster includes TPA and glyphosate, where majority of the altered protein expression was recorded and the other cluster includes DMBA and control having comparatively low number of altered proteins (Fig. 7). Moreover, calyculin and calgranulin-B were present only in the cluster of glyphosate and TPA whereas SOD 1 is higher in DMBA and control cluster in comparison to other cluster.

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The commercial formulation of the herbicide glyphosate (N-phosphonomethyl-glycine) Roundup Original® (glyphosate 41%, POEA≈15%—Monsanto Company, St. Louis, MO, USA) was used, which contains 360 g/l glyphosate acid equivalent

52 µg/mouse followed 1 week later by thrice a week application of TPA, 5 µg/mouse).

Group IV Glyphosate (s)+TPA (Single topical application of glyphosate, 25 mg/kg b.wt followed 1 week later by TPA application as in group III).

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Group VI DMBA (Single topical application of DMBA, 52 µg/mouse).

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Male Swiss albino mice (23–25 g body weight [bw]) were taken from Indian Institute of Toxicology Research (IITR) animal breeding colony and acclimatized for 1 week. The ethical approval for the experiment was obtained from institutional ethical committee. The animals were kept under standard laboratory conditions (temperature 23 ± 2 °C, relative humidity 55–58%) and were fed with synthetic pellet basal diet (Ashirwad, Chandigarh, India) and tap water ad libitum. Animals were randomly divided into 8 groups of 20 animals each. Hair were clipped in the dorsal region with proper care in an area of 2 cm² using electrical clippers, not lubricated with oil or grease. The long term treatment was given as described earlier [24]. Briefly,

- Group I: Untreated control (No treatment).
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- Group III: DMBA + TPA (Single topical application of DMBA, 52 µg/mouse followed 1 week later by thrice a week application of TPA, 5 µg/mouse).
- Group IV: Glyphosate (25 mg/kg bw) followed 1 week later by TPA application as in group III.
- Group V: Glyphosate (25 mg/kg bw) followed 1 week later by TPA application as in group III.
- Group VI: DMBA (Single topical application of DMBA, 52 µg/mouse).
- Group VII: TPA (Thrice a week topical application of TPA, 5 µg/mouse).
- Group VIII: DMBA + glyphosate (Single topical application of DMBA [52 µg in group III], followed 1 week later by topical treatment of glyphosate, 25 mg/kg bw thrice per week).

Vehicle for glyphosate, DMBA and TPA were 50% ethanol and acetone respectively.

Animals from all the groups were examined every week for gross morphological changes including body weight changes, development and volume of squamous cell papilloma (tumors) locally on the skin during the entire study period, and tumors larger than 1 mm diameter, were included in the total number of tumors. Tumor volume per tumor bearing mouse was calculated in each group using formula $V = D_1 \times D_2^2 \times \pi / 6$ (where D_1 = bigger dimension and D_2 = smaller dimension). All the surviving animals were sacrificed at the end of the study period, i.e. 32 weeks for complete carcinogenic, tumor initiating and promoting studies.

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I	Untreated	0/20	—	—	—	—	—
II	Glyphosate	0/20	—	—	—	—	—
III	DMBA + TPA	22/20 ^a	53	100	154	7.0 ± 1.1	36.4 ± 5.1
IV	Glyphosate (25 mg/kg bw) + TPA	0/20	—	—	—	—	—
V	Glyphosate (25 mg/kg bw) + DMBA	0/20	—	—	—	—	—
VI	DMBA	0/20	—	—	—	—	—
VII	TPA	0/20	—	—	—	—	—
VIII	DMBA + glyphosate	8/20 ^b	130	60	73	2.8 ± 0.9	26.2 ± 4.8

^a 2-DE, versus untreated group (ANOVA test, 4-stage dose, n = multiple dose). Details of treatment are provided in Materials and Methods section.

number of tumors was 7.8 ± 1.1 in group III. However, in group VII, it was 2.8 ± 0.9 (Fig. 1C, Table 1). These tumors were initiated as a minute wart like growth, which progressed during the course of experiment and average tumor volume was 36.4 ± 5.1 mm³ in group III and 26.2 ± 4.8 mm³ in group VIII (Fig. 1D, Table 1). These results clearly indicate significant tumor promoting potential of glyphosate in mouse skin model of carcinogenesis.

3.2. Protein expression profile

Using 2-DE, comparisons of differentially expressed proteins were made in mouse skin following topical treatment with glyphosate (50 mg/kg bw/mouse), TPA (10 µg/mouse) and DMBA (104 µg/mouse) with untreated mouse skin individually, using PDQuest 7.0.0 2-D gel analysis software. Representative 2-DE maps are shown in Fig. 2. Image matching derived from 4 groups showed a total of ~2600 spots. Out of these, 22 spots were differentially expressed, exhibiting > 2 fold change between values of treated and control animals (Fig. 2). These spots were excised from the gels and analyzed using MALDI-TOF/MS mass spectrometer. PMF from the proteins was obtained and the resulting spectra were used to identify the proteins with the Mascot search program. Protein spots that appeared more than once, were considered as the same protein and assigned the same number. These identified proteins were categorized according to their molecular functions (Table 2), biological functions and subcellular localization (Fig. 3A and B) as referred to SWISS-PROT database. Protein spots nos. 7 and 18–1, 18–2 were up-regulated and spot no. 13 was down-regulated by glyphosate and TPA treatment (Fig. 4). Related fingerprint mass spectra of calyculin, calgranulin-B and SOD are shown in Fig. 5.

3.3. Protein expression profile in glyphosate and TPA-treated mouse skin

Substantially common and differentially expressed protein spots among glyphosate and TPA-treated skin tissues were quantitatively analyzed individually. Comparison between the gels of glyphosate and TPA revealed that 13 specific proteins spots (1, 2, 3, 6, 1, 6, 2, 7, 8, 11, 12, 13, 15, 18–1, and 18–2)

from a total of 22 spots were showing the similar expression pattern. Among the selected and identified proteins with statistically significant altered expression (p < 0.05), we focused on the proteins involved in apoptosis and growth-inhibition, anti-oxidation, energy metabolism, angiogenesis, calcium binding and protein biosynthesis processes. These proteins are translation elongation factor eIF1 α alpha chain (eIF1A), catalase, ankyrin II (CA II), annexin II, cyclophilin, fibronectin, alpha-VIC1 antibody, peroxiredoxin 2 (PRX II), superoxide dismutase [Cu-Zn] (SOD 1), stefin A, and calgranulin-B (Fig. 6, Table 3).

3.4. Protein expression profile in glyphosate and DMBA treated mouse skin

Among the 22 differentially expressed protein spots, 4 specific spots (1, 3, 7, 1, and 12) were showing the similar expression pattern between the gels of glyphosate and DMBA treated skin tissues. These proteins are eIF1A, CA II, fibronectin, alpha-VIC1 antibody and PRX II (Fig. 6, Table 3).

3.5. Cluster analysis of differentially expressed proteins in control, glyphosate, TPA and DMBA treated mouse skin

To understand the carcinogenic activity of glyphosate in mouse skin based on the level of protein expression information generated on 2-DE gels hierarchical cluster analysis was applied. The analysis facilitated the visualization of groupings based on the protein expression changes, potentially showing the relationship between glyphosate and TPA, which further leads support to their tumor promoting activity. A hierarchical clustering map is generated with the differentially expressed protein spots. The analysis showed 2 major clusters, one cluster includes TPA and glyphosate, where majority of the altered protein expression was recorded and the other cluster includes DMBA and control having comparatively low number of altered proteins (Fig. 7). Moreover, calyculin and calgranulin-B were present only in the cluster of glyphosate and TPA whereas SOD 1 is higher in DMBA and control cluster in comparison to other cluster.

Table 1 – Role of carcinogenic, tumor initiating and tumor promoting effect of glyphosate in mouse skin model of carcinogenesis.

Groups	Treatment	Number of animals with tumors	1st induction of tumor (in days)	% of animals with tumors	Total number of tumors	Avg. no. of tumors/mouse (mean±SD)	Avg. tumor volume/tumor bearing mouse (mm ³) (mean±SD)
I	Untreated	0/20	–	–	–	–	–
II	Glyphosate	0/20	–	–	–	–	–
III	DMBA+TPA	20/20	52	100	156	7.8±1.1	96.4±5.1
IV	Glyphosate (s)+TPA	0/20	–	–	–	–	–
V	Glyphosate (m)+TPA	0/20	–	–	–	–	–
VI	DMBA (s)	0/20	–	–	–	–	–
VII	TPA	0/20	–	–	–	–	–
VIII	DMBA+glyphosate	8/20	130	40	23	2.8±0.9	26.2±4.8

* $p < 0.05$ versus untreated group (ANOVA test). s=single dose, m= multiple dose. Details of treatment are provided in Materials and methods section.

number of tumors was 7.8 ± 1.1 in group III, however, in group VIII, it was 2.8 ± 0.9 (Fig. 1C; Table 1). These tumors were initiated as a minute wart like growth, which progressed during the course of experiment and average tumor volume was $96.4 \pm 5.1 \text{ mm}^3$ in group III and $26.2 \pm 4.8 \text{ mm}^3$ in group VIII (Fig. 1D; Table 1). These results clearly indicate significant tumor promoting potential of glyphosate in mouse skin model of carcinogenesis.

3.2. Protein expression profile

Using 2-DE, comparisons of differentially expressed proteins

from a total of 22 spots were showing the similar expression pattern. Among the selected and identified proteins with statistically significant altered expression ($p < 0.05$), we focused on the proteins involved in apoptosis and growth-inhibition, anti-oxidation, energy metabolism, angiogenesis, calcium binding and protein biosynthesis processes. These proteins are translation elongation factor eEF-1 alpha chain (eEF1A1), carbonic anhydrase 3 (CA III), annexin II, calcyclin, fab fragment of anti-VEGF antibody, peroxiredoxin-2 (PRX II), superoxide dismutase [Cu-Zn] (SOD 1), stefin A3 and calgranulin-B (Fig. 6, Table 3).

Three Pillars of Cancer Science

Toxicology



Studies on glyphosate-induced carcinogenicity in mouse skin: A proteomic approach

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ABSTRACT

Glyphosate is a widely used broad spectrum herbicide, reported to induce various toxic effects in non-target species, but its carcinogenic potential is still unknown. Here we showed the carcinogenic effects of glyphosate using 2-stage mouse skin carcinogenesis model and proteomic analysis. Carcinogenicity study revealed that glyphosate has tumor promoting activity. Proteomic analysis using 2-dimensional gel electrophoresis and mass spectrometry showed that 22 spots were differentially expressed (>2 fold) on glyphosate, 7, 12-dimethylbenzylisothiazolone (DMBA) and 12-O-tetradecanoylphorbol-13-acetate (TPA) application over untreated control. Among them, 9 proteins (translational elongation factor eIF1 α , alpha chain, catalase, ankyrin II, annexin II, cyclophilin, fibronectin, alpha-VIC1 antibody, pectatelectin-2, superoxide dismutase [Cu-Zn], stefin A), and calgranulin-B were common and showed similar expression pattern in glyphosate and TPA-treated mouse skin. These proteins are known to be involved in several key processes like apoptosis and growth-inhibition, anti-oxidant responses, etc. The up-regulation of cyclophilin, calgranulin-B and down-regulation of superoxide dismutase [Cu-Zn] was further confirmed by immunoblotting, indicating that these proteins can be good candidate biomarkers for skin carcinogenesis induced by glyphosate. Altogether, these results suggested that glyphosate has tumor promoting potential in skin carcinogenesis and its mechanism seems to be similar to TPA.

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1. Introduction

Herbicides, used extensively for controlling pest and destroying weeds are ubiquitous contaminants accumulating in environment and hence humans get unavoidably exposed to these pesticides. About 3 billion tons of pesticides are used every year, on agricultural crops worldwide [1]. In some cases, even short term exposure of the pesticides can make impact on human health. Apart from the other toxic effects, pesticides are reported to cause genotoxicity/carcinogenicity also. Some pesticides have been classified as carcinogens by the United States Environmental Protection Agency (USEPA) [2] and International Agency for Research on Cancer (IARC) [3]. Glyphosate, N-(phosphonomethyl) glycine, commonly sold as

a commercial formulation named, Roundup is a widely used herbicide on both cropland and non-cropland areas [4]. The potential activity of glyphosate is through competitive inhibition of the *5-enolpyruvyl-shikimate-phosphate* synthase, an enzyme essential to the synthesis of aromatic amino acids in plants [5]. Toxicological profile of glyphosate, showed that it is a comparatively safe herbicide for animals [6]. Glyphosate shows also with its formulation products, such as surfactants and permeabilizing agents is usually considered to be harmless under both normal usage and chronic exposure [4]. In 1991, USEPA categorized this compound into class I, which means that it is probably not carcinogenic to humans [7]. Despite these reports, some case-control studies suggested an association between glyphosate exposure and the risk of non-

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Hodgkin's lymphoma [8,9]. In another study, both technical grade glyphosate and Roundup were shown to cause a rapid increase in cell division in human breast cancer cells [10]. Glyphosate has also been shown as a skin irritant [11]. Regarding the genotoxic potential, glyphosate exposure to human lymphocytes *in vitro* resulted in increased sister chromatid exchanges [12], chromosomal aberrations [13], and indicators of oxidative stress [14]. A recent study from our laboratory also showed the clastogenic effects of glyphosate in bone marrow cells of Swiss albino mice [15]. These reports prompted us to investigate its carcinogenic effect in long-term animal bioassay.

To evaluate toxicity/carcinogenicity induced by physical and chemical agents including pesticides, various test systems have been employed in bacteria, rodents and mammalian cells [16–18]. Each of these environmental challenges causes changes in DNA conformation, alterations in the levels of mRNA and protein expression, and post-translational modifications such as phosphorylation and glycosylation of proteins specific to each stressor [19]. In recent years, there has been considerable interest in linking carcinogenic/oxidative responses to gene and protein expression. Toxicoproteomics has received a lot of attention as a valuable tool to search reliable early predictive toxicity markers in response to environmental stimuli [20]. Two-dimensional gel electrophoresis (2-DE) coupled with mass spectrometry (MS), a high-throughput technique allows proteins of interest to be identified by their expression and/or modification pattern rather than using the traditional approach of translating gene expression data. Biomarkers can be used to identify causal associations and to make better quantitative estimates of these associations at relevant levels of exposure [21]. Yamamoto et al. [22] have utilized proteomic approach to identify potential biomarker candidates of hepatocellular carcinoma in rat liver.

Skin is the largest organ in the body and dermal contact is one of the most probable routes of human exposure to pesticides. Thus, mouse skin model represents a logical experimental choice [23]. As the long-term bioassay for carcinogenicity is expensive, time consuming and involves a large number of animals and ethical issues, development of biomarkers after short-term exposure are needed. The present investigation was carried out to study the carcinogenic potential of glyphosate and to identify differentially expressed proteins, using 2-DE and MS analysis after treatment with glyphosate, a known tumor promoter, 12-O-tetradecanoylphorbol-13-acetate (TPA) and tumor initiator, 7, 12-dimethylbenzylisothiazolone (DMBA) in mouse skin. Altered proteins identified through proteomic approach in our study may be potentially useful as early biomarkers, to detect the adverse effects of glyphosate.

2. Materials and methods

2.1. Materials

The commercial formulation of the herbicide glyphosate (N-phosphonomethyl glycine) Roundup Original[®] (glyphosate 41%, POEA +15%–Monsanto Company, St. Louis, MO, USA) was used, which contains 360 g/l glyphosate acid equivalent

as the isopropylamine salt and was procured from local market. Immobilized pH gradient (IPG) strips and 0.5% pH3–10/IC buffer were purchased from Bio-Rad Laboratories (Hercules, CA, USA). DMBA, TPA, CHAPS, DTT, and beta-actin (clone AC-20) antibody were from Sigma-Aldrich (Missouri, USA). DNase/RNase were from Bangalore Genei (Bangalore, India). The rest of the chemicals used in the study were of analytical grade of purity and procured locally.

2.2. Animals and treatments

2.2.1. Carcinogenicity study
Male Swiss albino mice (23–25 g body weight [bw]) were taken from Indian Institute of Toxicology Research (IITR) animal breeding colony and acclimatized for 1 week. The ethical approval for the experiment was obtained from institutional ethical committee. The animals were kept under standard laboratory conditions (temperature 23 ± 2 °C, relative humidity 55–55%) and were fed with synthetic pellet basal diet (Ashirwad, Chandigarh, India) and tap water ad libitum. Animals were randomly divided into 8 groups of 20 animals each. Hair were clipped in the dorsal region with proper care in an area of 2 cm² using electrical clippers, not lubricated with oil or grease. The long term treatment was given as described earlier [24]. Briefly,

- Group I: Untreated control (No treatment).
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- Group III: DMBA + TPA (Single topical application of DMBA, 52 µg/mouse followed 1 week later by thrice a week application of TPA, 5 µg/mouse).
- Group IV: Glyphosate (a) + TPA (Single topical application of glyphosate, 25 mg/kg bw followed 1 week later by TPA application as in group III).
- Group V: Glyphosate (a) + TPA (Thrice a week topical application of glyphosate, 25 mg/kg bw for 3 weeks [total 9 applications], followed 1 week later by TPA application as in group III).
- Group VI: DMBA (Single topical application of DMBA, 52 µg/mouse).
- Group VII: TPA (Thrice a week topical application of TPA, 5 µg/mouse).
- Group VIII: DMBA + glyphosate (Single topical application of DMBA [a] in group III, followed 1 week later by topical treatment of glyphosate, 25 mg/kg bw thrice per week).

Vehicle for glyphosate, DMBA and TPA were 50% ethanol and acetone respectively.

Animals from all the groups were examined every week for gross morphological changes including body weight changes, development and volume of squamous cell papilloma (tumors) locally on the skin during the entire study period, and tumors larger than 1 mm diameter, were included in the total number of tumors. Tumor volume per tumor bearing mouse was calculated in each group using formula $V = D_1^2 \times D_2^2 \times \pi$ (where D_1 = bigger dimension and D_2 = smaller dimension). All the surviving animals were sacrificed at the end of the study period, i.e. 32 weeks for complete carcinogenic, tumor initiating and promoting studies.

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V	Glyphosate (a) + TPA	0/20	–	–	–	–	–
VI	DMBA (b)	0/20	–	–	–	–	–
VII	TPA	0/20	–	–	–	–	–
VIII	DMBA + glyphosate	8/20	130	60	73	2.8 ± 0.9	26.2 ± 4.8

T = 0 h; DMBA, vehicle untreated group (ANOVA test); a = single dose, b = multiple dose. Details of treatment are provided in Materials and Methods section.

number of tumors was 7.8 ± 1.1 in group III. However, in group VII, it was 2.8 ± 0.9 (Fig. 1C, Table 1). These tumors were initiated as a minute wart like growth, which progressed during the course of experiment and average tumor volume was 36.4 ± 5.1 mm³ in group III and 26.2 ± 4.8 mm³ in group VIII (Fig. 1D, Table 1). These results clearly indicate significant tumor promoting potential of glyphosate in mouse skin model of carcinogenesis.

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Using 2-DE, comparisons of differentially expressed proteins were made in mouse skin following topical treatment with glyphosate (50 mg/kg bw/mouse), TPA (10 µg/mouse) and DMBA (104 µg/mouse) with untreated mouse skin individually, using PDQuest 7.0.0 2-D gel analysis software. Representative 2-DE maps are shown in Fig. 2. Image matching derived from 4 groups showed a total of ~2600 spots. Out of these, 22 spots were differentially expressed, exhibiting > 2 fold change between values of treated and control animals (Fig. 2). These spots were excised from the gels and analyzed using MALDI-TOF/MS mass spectrometer. PMF from the proteins was obtained and the resulting spectra were used to identify the proteins with the Mascot search program. Protein spots that appeared more than once, were considered as the same protein and assigned the same number. These identified proteins were categorized according to their molecular functions (Table 2), biological functions and subcellular localization (Fig. 3A and B) as referred to SWISS-PROT database. Protein spots nos. 7 and 18–1, 18–2 were up-regulated and spot no. 13 was down-regulated by glyphosate and TPA treatment (Fig. 4). Related fingerprint mass spectra of calyculin, calgranulin-B and SOD are shown in Fig. 5.

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Substantially common and differentially expressed protein spots among glyphosate and TPA-treated skin tissues were quantitatively analyzed individually. Comparison between the gels of glyphosate and TPA revealed that 13 specific proteins spots (1, 2, 3, 6, 1, 6, 2, 7, 8, 11, 12, 13, 15, 18–1, and 18–2)

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Three Pillars of Cancer Science

Toxicology

From: HEYDENS, WILLIAM F [AG/1000]
Sent: Thursday, August 06, 2015 9:55 AM
To: 'Ashley Roberts Intertek'; FARMER, DONNA R [AG/1000]
Subject: RE: Keith

Ashley,
I think the short answer is no. The focus of this is what is the carcinogenic potential of glyphosate.

That said, the surfactant in the formulation will come up in the tumor promotion skin study because we think it played a role there.

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He has asked if we need to give any consideration to exposures of formulants in the commercial product, at least in applicators? I was under the impression these were inert but reading a response this morning in the Ecologist makes it sound like it is the combination that is toxic!!!

What do you think?

Plaintiff Exhibit
0366

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Three Pillars of Cancer Science

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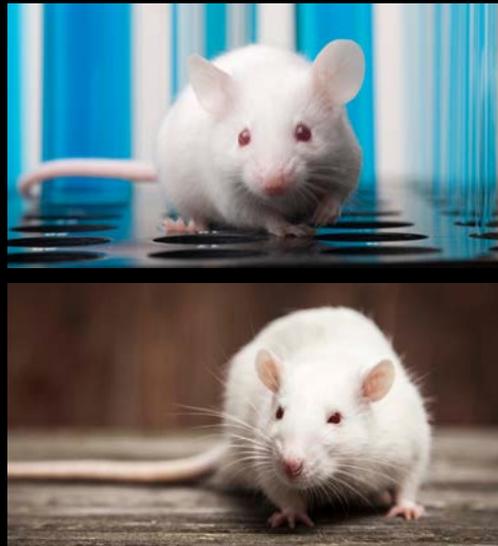
Epidemiology



IARC Classification

1. Sufficient
2. Limited
3. Inadequate
4. Lack of Carcinogenicity

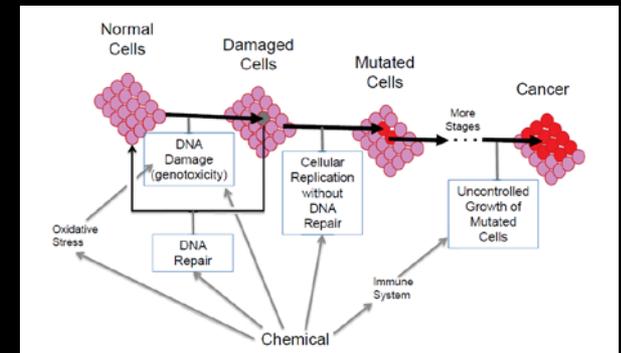
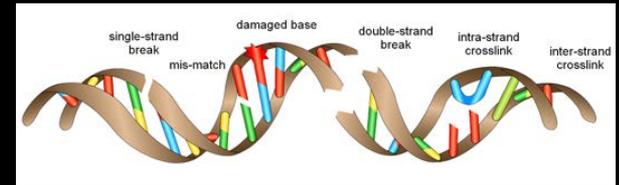
Toxicology



IARC Classification

1. Sufficient
2. Limited
3. Inadequate
4. Lack of Carcinogenicity

Mechanism



IARC Classification

1. Strong
2. Moderate
3. Weak

3 Core Questions

YES

1. Can Roundup be a substantial contributing factor in causing cancer?
2. Was Roundup a substantial contributing factor in causing Mr. Johnson's cancer?
3. Did Monsanto act with knowing disregard of human health?

2. Was Roundup a substantial contributing factor in causing Mr. Johnson's cancer?

Jury Instruction

A substantial factor in causing harm is a factor that a reasonable person would consider to have contributed to the harm. It must be more than a remote or trivial factor. It does not have to be the only cause of the harm. Conduct is not a substantial factor in causing harm if the same harm would have occurred without that conduct.

Mr. Johnson's Cancer

Jun.
2012

Mr. Johnson is promoted to integrated pest manager for Benicia School District



Mr. Johnson's Cancer

Full spraying season
50 gallons / hour

Jun.
2012

Mr. Johnson is promoted
to integrated pest
manager for Benicia
School District



Aug.
2012

Mr. Johnson completes
first spraying season
using truck sprayer



Mr. Johnson's Cancer

Full spraying season
50 gallons / hour

Intermittent
spraying

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Mr. Johnson's Cancer

May.
2013

Mr. Johnson begins
second intensive spraying
season



Mr. Johnson's Cancer

Summer
2013

Mr. Johnson gets massive exposure during Mary Farmer incident



May.
2013

Mr. Johnson begins second intensive spraying season



Mr. Johnson's Cancer

Summer
2013

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May.
2013

Mr. Johnson begins second intensive spraying season



Aug.
2013

Mr. Johnson completes second season of spraying



Mr. Johnson's Cancer

Summer
2013

Mr. Johnson gets massive exposure during Mary Farmer incident



May.
2013

Mr. Johnson begins second intensive spraying season



Aug.
2013

Mr. Johnson completes second season of spraying



Mr. Johnson's Cancer



Sep.
2013

Mr. Johnson presents to doctor with several wasp stings. Medical record at the time states “**Negative for rash**” Tr. 3131:8.

Mr. Johnson's Cancer

Dec.
2013

Mr. Johnson presents for lumbar injury, medical record states "no deformity, no shift, no scarring, no swelling." Tr. 3232:16-17

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Mr. Johnson presents to doctor with several wasp stings. Medical record at the time states "Negative for rash" Tr. 3131:8.

Mr. Johnson's Cancer

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Mr. Johnson's Cancer

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Mr. Johnson's Cancer



May
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Mr. Johnson develops a rash on his skin that does not respond to normal treatment.

Mr. Johnson's Cancer

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Following several biopsies, Mr. Johnson is diagnosed with mycosis fungoides.

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Mr. Johnson's Cancer

Oct. 2014



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Mr. Johnson's Cancer



Oct.
2014

IARC announces
investigation into
glyphosate



Message

From: HEYDENS, WILLIAM F [AG/1000] [REDACTED]@monsanto.com
Sent: 10/15/2014 9:08:37 PM
To: [REDACTED]@monsanto.com
CC: [REDACTED]@monsanto.com; FARMER, DONNA R [AG/1000]
[REDACTED]@monsanto.com]; SALTIRAS, DAVID A [AG/1000] [REDACTED]@monsanto.com]; KOCH,
MICHAEL S [AG/1000] [REDACTED]@monsanto.com
Subject: IARC Evaluation of Glyphosate

[REDACTED]

It is my recollection that you notified the EU-GTF of this IARC evaluation, but I am not aware that there has been any talk of approaching the GTF about providing funding to fight this because it is not considered in the remit of achieving Annex I renewal. If so, is this really the case? I thought the EU evaluation could go well into the summer of 2015, and wouldn't an adverse IARC evaluation have the real potential to impact the results of the Annex I renewal?

I really started thinking about this after our phone call yesterday with the outside epidemiology experts that Donna lined up. The bottom line of the call was that there really is no meaningful publication that we can complete prior to the February submission to positively impact the epidemiology discussion outcome in March. One has to consider that this situational timing did not happen by chance and that more than just pure bad luck is working against glyphosate.

And while we have vulnerability in the area of epidemiology, we also have potential vulnerabilities in the other areas that IARC will consider. namely. exposure. genotox. and mode of action (David has the animal onco studies under

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And while we have vulnerability in the area of epidemiology, we also have potential vulnerabilities in the other areas that IARC will consider, namely, exposure, genotox, and mode of action (David has the animal onco studies under control). If there is a force working against glyphosate, there is ample fodder to string together to help the cause even though it is not scientifically justified in its purest form. Putting all this in the proper perspective will be quite resource intensive, so can't we consider approaching the GTF? Recall that the PAG already agreed to fund the onco publication 2+ years ago for this exact reason.

Thanks.

Bill

Mr. Johnson's Cancer



Oct.
2014

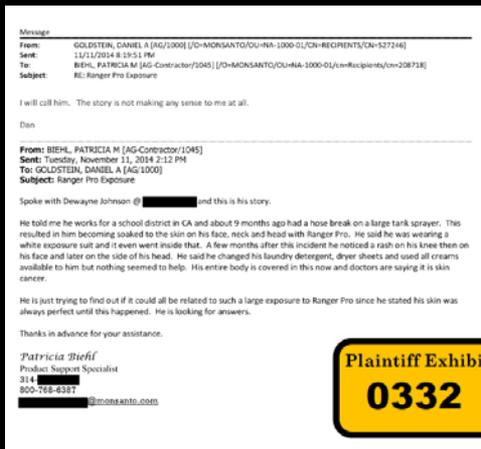
IARC announces
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Mr. Johnson's Cancer

Oct. 2014 IARC announces investigation into glyphosate

Nov. 2014 Mr. Johnson calls Monsanto looking for answers.



Plaintiff Exhibit
0332

Sent: 11/11/2014 8:19:51 PM
To: BIEHL, PATRICIA M [AG-Contractor/1045] [/O=MONSANTO/OU=NA-1000-01/cn=Recipients/cn=208718]
Subject: RE: Ranger Pro Exposure

I will call him. The story is not making any sense to me at all.

Dan

From: BIEHL, PATRICIA M [AG-Contractor/1045]
Sent: Tuesday, November 11, 2014 2:12 PM
To: GOLDSTEIN, DANIEL A [AG/1000]
Subject: Ranger Pro Exposure

Spoke with Dewayne Johnson @ [REDACTED] and this is his story.

He told me he works for a school district in CA and about 9 months ago had a hose break on a large tank sprayer. This resulted in him becoming soaked to the skin on his face, neck and head with Ranger Pro. He said he was wearing a white exposure suit and it even went inside that. A few months after this incident he noticed a rash on his knee then on his face and later on the side of his head. He said he changed his laundry detergent, dryer sheets and used all creams available to him but nothing seemed to help. His entire body is covered in this now and doctors are saying it is skin cancer.

He is just trying to find out if it could all be related to such a large exposure to Ranger Pro since he stated his skin was always perfect until this happened. He is looking for answers.

Thanks in advance for your assistance.

Patricia Biehl

Product Support Specialist

314-[REDACTED]

800-768-6387

[REDACTED]@monsanto.com

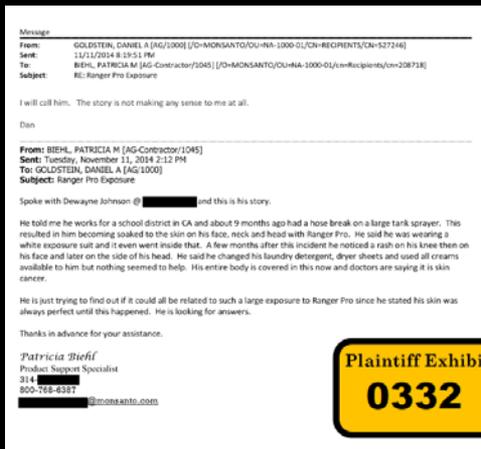
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Mr. Johnson's Cancer

Oct. 2014 IARC announces investigation into glyphosate

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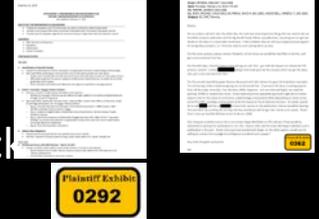


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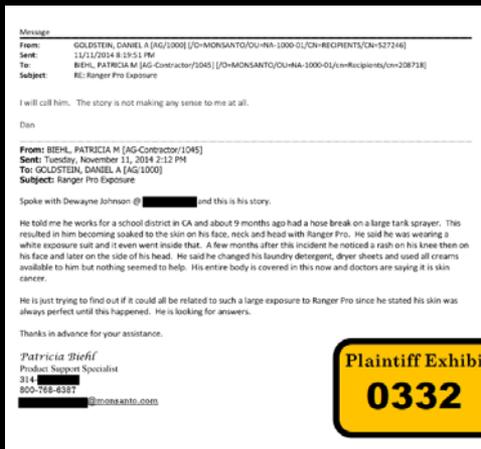
Feb. 2015 Monsanto plans to attack IARC ruling.



Mr. Johnson Continues to Spray

Mr. Johnson calls Monsanto looking for answers.

Nov. 2014



Plaintiff Exhibit
0332

From: HEYDENS, WILLIAM F [AG/1000]

Sent: Thursday, February 19, 2015 7:53 AM

To: FARMER, DONNA R [AG/1000]

Cc: KOCH, MICHAEL S [AG/1000]; SALTMIRAS, DAVID A [AG/1000]; HODGE-BELL, KIMBERLY C [AG/1000]

Subject: RE: IARC Planning

Donna,

Per our phone call with John the other day, the next two most important things that we need to do are the Meta-analysis publication and the Ag Health Study Follow-up publication, assuming we can get our hands on the data in a reasonable timeframe. I feel confident that we will have organizational support for doing these projects, so I think we need to start setting them up now.

For the meta-analysis, please contact Elizabeth, let her know we would like her/Ellen to do this, and get a cost estimate from her.

For the AHS data, I heard 2 action items during our call: first - get with the lawyers to initiate the FOI process; second - contact [REDACTED] and get him lined up to do the analysis when we get the data; also, get a cost estimate from him.

For the overall plausibility paper that we discussed with John (where he gave the butadiene example), I'm still having a little trouble wrapping my mind around that. If we went full-bore, involving experts from all the major areas (Epi, Tox, Genetox, MOA, Exposure - not sure who we'd get), we could be pushing \$250K or maybe even more. A less expensive/more palatable approach might be to involve

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One thing we could do now on this is to contact Roger McClellan at CRC and see if they would be amenable to putting this publication in *Crit. Rev. Toxicol.* John said he knew that Roger had done such a publication in the past. David, since you have worked with Roger on the other papers, would you be willing to contact him to judge his willingness to publish such a paper?

Any other thoughts welcomed.

Bill

Plaintiff Exhibit

0362

Political Science

- Unfortunately, we are facing regulatory reviews with increased focus on
 - Claims in the peer reviewed literature, irrespective of the quality of the science
 - Stakeholder input including activist researchers
 - Political pressure on outcomes – e.g. POEAs in Germany
 - Reduced pesticide use in general
- Williams et al. (2000) has served us well in toxicology over the last decade
- We need a stronger arsenal of robust scientific papers to support the safe use of our products as we face the next set of chemistry registration reviews across the globe
- With increasing business interests in South America, a local network credible expert scientists is crucial to facilitate scientifically robust and objective regulatory evaluations of our products *We have not determined exactly what we should & could do here. I would modify bullet to reflect that we need to determine an appropriate & do-able (i.e., we can get someone to pay for it course of action here*

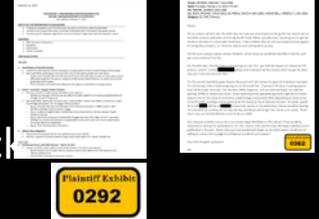
Plaintiff Exhibit

0373

Mr. Johnson's Cancer

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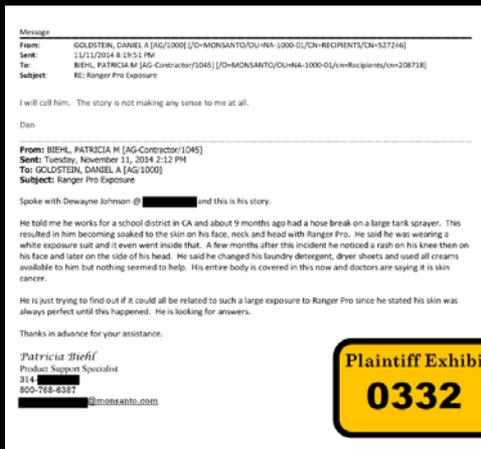
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Mr. Johnson Continues to Spray

Mr. Johnson calls Monsanto looking for answers.

Nov. 2014



Plaintiff Exhibit
0332

Draft Feb 23, 2015

Plaintiff Exhibit

0292

**ATTACHMENT A: PREPAREDNESS AND ENGAGEMENT PLAN
FOR IARC CARCINOGEN RATING OF GLYPHOSATE**

Last Updated: February 17, 2015

OBJECTIVES FOR PREPAREDNESS & ENGAGEMENT

- Protect the reputation and FTO of Roundup by communicating the safety of glyphosate
- Amplify science-based information to prevent unfounded claims from becoming popular opinion
- Provide cover for regulatory agencies to continue making re-registration decisions based on science

AUDIENCES

- IARC Panelists and Observers
- Regulators
- Stakeholders
- Farmer Customers

STRATEGIES/TACTICS

PRE-IARC

1. Amplification of Scientific Studies

- Support the development of three new papers on glyphosate focused on epidemiology and toxicology
- Work with RPSA and Strategic Communications to amplify existing studies and new papers
 - Authors work directly with scientific journals to issue alerts and news releases on new bodies of work
 - RPSA posts blog from first-person viewpoint of Monsanto's David Saltmiras, co-author of one of the

Mr. Johnson's Cancer

Draft Feb 23, 2015



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- Develop a toolkit containing key information and resources
- Identify any message shortcomings and address through updates to monsanto.com/glyphosate and through US and EU blog posts
- Work with RPSA, Stakeholder Outreach Team, Industry Affairs, Government Affairs, US Business, Global CE and Regulatory teams, etc. to engage industry partners
 - Tier 1: Crop Life International / European Crop Protection Association / GMO Answer identify committees that are best to engage
 - Tier 2: Academics (AgBioChatter), Biofortified, Sense About Science, Genetic Literacy Academics Review
 - Tier 3: Alert food companies via Stakeholder Engagement team (IFIC, GMA, CFI) for “strategy” to provide early education on glyphosate residue levels, describe science-based studies versus agenda-driven hypotheses
 - Tier 4: Inoculate key grower associations

Plaintiff Exhibit
0292

3. Address New Allegations

- Respond quickly and publically to new pseudoscience cancer studies
- Identify / request third-party experts to blog, op/ed, tweet and/or link, repost, retweet, etc.

POST-IARC

4. Orchestrate Outcry with IARC Decision ~ March 10, 2015

- Industry conducts robust media / social media outreach on process and outcome
 - [Sense About Science?] leads industry response and provides platform for IARC observers and industry spokesperson
 - CLI and other associations issue press releases

FROM: BIEHL, PATRICIA M (AG-CO/2009/1045)
Sent: Tuesday, November 11, 2014 2:12 PM
To: GOLDSTEIN, DANIEL A (AG 1000)
Subject: Ranger Pro Exposure

Spoke with Dewayne Johnson @ [REDACTED] and this is his story.

He told me he works for a school district in CA and about 9 months ago had a hose break on a large tank sprayer. This resulted in him becoming soaked to the skin on his face, neck and head with Ranger Pro. He said he was wearing a white exposure suit and it even went inside that. A few months after this incident he noticed a rash on his knee then on his face and later on the side of his head. He said he changed his laundry detergent, dryer sheets and used all creams available to him but nothing seemed to help. His entire body is covered in this now and doctors are saying it is skin cancer.

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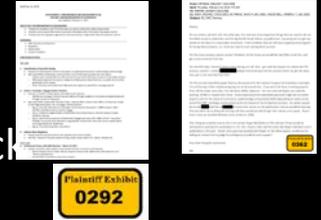
Patricia Biehl
Product Support Specialist
314 [REDACTED]
800-710-6127 | pbiehl@monsanto.com

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Mr. Johnson's Cancer

Oct. 2014 IARC announces investigation into glyphosate

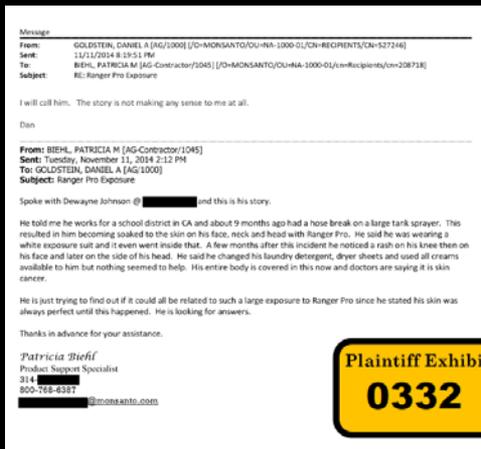
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Mr. Johnson Continues to Spray

Mr. Johnson calls Monsanto looking for answers.

Nov. 2014



Plaintiff Exhibit
0332

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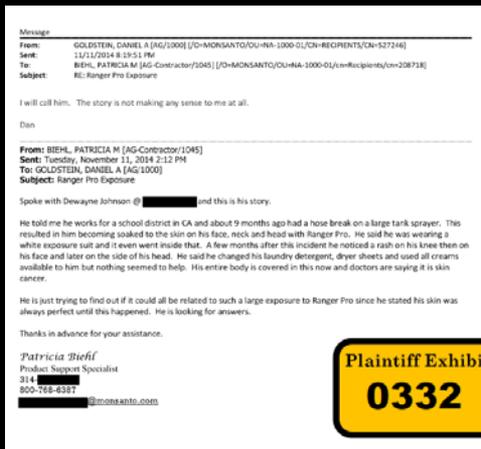


Plaintiff Exhibit
0292

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Nov. 2014 Mr. Johnson calls Monsanto looking for answers.

Mar. 2015 IARC classifies glyphosate as Class 2A carcinogen



Plaintiff Exhibit
0332



Mr. Johnson's Cancer

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Plaintiff Exhibit
0292

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Nov. 2014

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Mar. 2015

IARC classifies glyphosate as Class 2A carcinogen

Message

From: GOLDSTEIN, DANIEL A [AG 1000] [/O=MONSANTO/OU=NA-1000-01/CN=REGENTS/CN=527246]
Sent: 11/11/2014 10:25:11 AM
To: BIEHL, PATRICIA M [AG-Contractor/1045] [/O=MONSANTO/OU=NA-1000-01/CN=Recipients/cm=208718]
Subject: RE: Ranger Pro Exposure

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Patricia Biehl
Product Support Specialist
314 [REDACTED]
800-760-6327 @monsanto.com

Plaintiff Exhibit
0332



Mr. Johnson's Cancer

Mar.
2015

Mr. Johnson reaches out to Missouri Regional Poison Control, seeking answers.



Human Exposure / Adverse Effect Incidents Involving Monsanto Agricultural Products

Reporting Categories: H-A, H-B, H-C

Reporting Period: March 1, 2015 – March 31, 2015

Substance:	Ranger Pro Herbicide from Monsanto
Serial Number:	32283189
Date:	03/27/2015
Medical Outcome:	Major Effect H-B
EPA Reg. No.	524-517
Active Ingredients:	Glyphosate 41%
State:	California
History and Notes:	Caller states he has been using Ranger Pro as part of his job for 2 to 3 years. He has recently been diagnosed with cutaneous T cell lymphoma. He has concerns about continuing to use Roundup as part of his job and questions if Roundup could be a source of his cancer. As the call progressed, caller said that doctors are unsure as to how to treat his condition and they are not even sure if it is cancer. Caller states that he works with Ranger Pro using a 50 gallon tank and also using a backpack sprayer. He dilutes 10 ounces

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State:	California
History and Notes:	<p>Caller states he has been using Ranger Pro as part of his job for 2 to 3 years. He has recently been diagnosed with cutaneous T cell lymphoma. He has concerns about continuing to use Roundup as part of his job and questions if Roundup could be a source of his cancer. As the call progressed, caller said that doctors are unsure as to how to treat his condition and they are not even sure if it is cancer. Caller states that he works with Ranger Pro using a 50 gallon tank and also using a backpack sprayer. He dilutes 10 ounces of the Roundup per gallon (3.0%) for the 50 gallon tank and 4 ounces of Roundup per gallon (1.25%) when using the backpack sprayer. He recalls having been exposed to Roundup twice in the past 2 to 3 years, both from the backpack leaking/malfunctioning. In one case, he was wearing personal protective equipment (PPE) but it soaked through the PPE and his clothing. Recently, he has had a swollen foot and the MD's cannot figure out what is going on. The caller's level of fear is rising over his continued use of Ranger Pro. He states he continues to get unexplained rashes and nodules over his body. MRPC discussed the product toxicity. The symptoms are not an expected response from the product. Advised MRPC is available, if the treating MD has any questions.</p>

Mr. Johnson's Cancer

Mar.
2015

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Mr. Johnson's Cancer

Mar. 2015



Mr. Johnson's Cancer

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Apr.
2015

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Mr. Johnson's Cancer

Mar.
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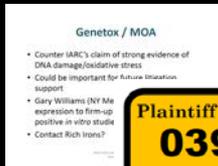
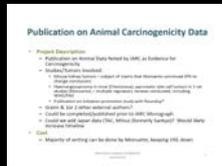
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May
2015

Monsanto raises litigation concerns.



Proposal for Post-IARC Meeting Scientific Projects

DRAFT

May 11, 2015

Mr. Johnson's Cancer

Mar.
2015

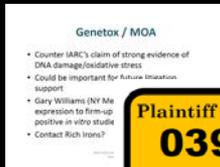
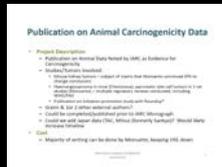
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Monsanto raises litigation concerns.



Why do more?

- Severe stigma attached to Group 2A Classification
- Aaron Blair continues to defend work & exaggerate number of studies w/ association while ignoring AHS
- In response to our critique, can expect IARC to beef-up monograph as much as possible
- IARC plans to pool data globally in the future
 - Blair announced at meeting that he has already put together an unofficial work group to begin the process
 - North American Pooled Project (NAPP) already underway and early results reported in 2014
 - Believe this will be used to move pesticides to Group 1
- Provide additional support ('air cover') for future regulatory reviews
 - Broad EU review recently recommended by BfR
 - Other regulatory agencies stated they will review after Monograph publishes
- ASTDR evaluation
- [REDACTED]
- Litigation support

Mr. Johnson's Cancer

Mar.
2015

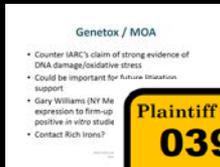
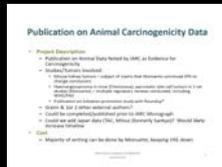
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Genetox / MOA

- Counter IARC's claim of strong evidence of DNA damage/oxidative stress
- Could be important for future litigation support
- Gary Williams (NY Medical College) - Use gene expression to firm-up non-genotoxic MOA in positive *in vitro* studies with formulations
- Contact Rich Irons?

Mr. Johnson's Cancer

Mar.
2015

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Mr. Johnson Continues to Spray

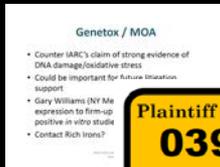
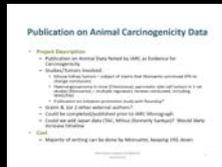
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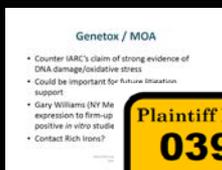
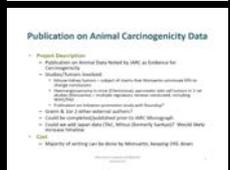
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Sept.
2015

Monsanto raises concern that IARC finding will reduce sales in California school districts.



May
2015

Monsanto raises litigation concerns.



Wilbur-Ellis Company

1427 Abbott Street

Salinas, CA 93901

Cell: (408) [REDACTED]

Fax: (650) [REDACTED]

[REDACTED]@wilburellis.com

-----Original Message-----

From: GOULD, STEVEN D [AG/1000] [mailto:[REDACTED]@monsanto.com]

Sent: Thursday, September 10, 2015 1:08 PM

To: Greg Fernald

Subject: RE: [REDACTED] Moves to Label Monsanto's Roundup 'Carcinogenic' | [REDACTED]

He sent to me too. It's hard to understand how against all science and law the can do this.

Steve Gould

Steven D. Gould

Monsanto IT&O

Account Manager

951-894-[REDACTED] office

951-704-[REDACTED] mobile

monsantoITO.com

-----Original Message-----

From: Greg Fernald [mailto:[REDACTED]@wilburellis.com]

To: HARDY, JOHN O [AG/1000] [/O=MONSANTO/OU=NA-1000-01/cn=Recipients/cn=53364]
Subject: Fwd: [REDACTED] Moves to Label Monsanto's Roundup 'Carcinogenic' | [REDACTED]

I liked this analogy from Greg

Steve Gould

Begin forwarded message:

From: Greg Fernald <[REDACTED]@wilburellis.com>
Date: September 10, 2015 at 3:19:21 PM PDT
To: "GOULD, STEVEN D [AG/1000]" <steven.d.gould@monsanto.com> [REDACTED]
Subject: RE: [REDACTED] Moves to Label Monsanto's Roundup 'Carcinogenic' | [REDACTED]

We are being overrun by liberals and morons...sort of like a zombie movie, so we just have to start taking them out one at a time, starting with the elections next year.

Greg Fernald
Professional Markets

Wilbur-Ellis Company
1427 Abbott Street
Salinas, CA 93901
Cell: (408) [REDACTED]
Fax: (650) [REDACTED]
[REDACTED]@wilburellis.com

-----Original Message-----

From: GOULD, STEVEN D [AG/1000] [mailto:[REDACTED]@monsanto.com]
Sent: Thursday, September 10, 2015 1:08 PM

Mr. Johnson's Cancer

Message

From: GOULD, STEVEN D [AG/1000] [/O=MONSANTO/OU=NA-1000-01/CN=RECIPIENTS/CN=116457]
Sent: 9/10/2015 10:41:00 PM
To: HARDY, JOHN O [AG/1000] [/O=MONSANTO/OU=NA-1000-01/cn=Recipients/cn=53364]
Subject: Fwd: [REDACTED] Moves to Label Monsanto's Roundup 'Carcinogenic' | [REDACTED]

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Mr. Johnson's Cancer

Mar.
2015

Mr. Johnson reaches out to Missouri Regional Poison Control, seeking answers.



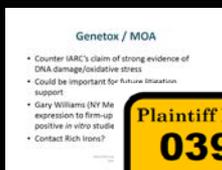
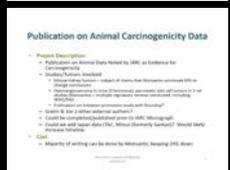
Mr. Johnson Continues to Spray

Apr.
2015

Dr. Ofodile writes letter to Benicia asking for Mr. Johnson to stop spraying chemicals.

Sept.
2015

Monsanto raises concern that IARC finding will reduce sales in California school districts.



May
2015

Monsanto raises litigation concerns.



cost on CA Municipalities Markets

By Steve Gould

Volumes:

I would estimate a gradual 2-3 year loss of up to 200 K REGS of all tiers of Roundup IT&O Glyphosate.

High Tier 65K

Mid Tier 75 K

Low Tier 60K

The volume will come from customers like CALTRANS. Caltrans Volume is a mix of high and mid tier and we stand to loss 5-10K depending on internal decisions

School districts are another big risk with the healthy schools act and increased attention. They frequently use PROMAX and PRO Concentrate today.

Airports, arenas, stadiums, municipal buildings any public facility especially if it goes out to bid could be affected

Cities, Counties, reservoirs, wildlife management areas and other similar locations could also be affected.

We will not really know the costs until we see how the agencies react and how many of them must

Low Tier 60K

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Airports, arenas, stadiums, municipal buildings any public facility especially if it goes out to bid could be affected

Cities, Counties, reservoirs, wildlife management areas and other similar locations could also be affected.

We will not really know the costs until we see how the agencies react and how many of them must address this.

This volume is based on discussing with distributor representatives, Monsanto sales volume estimates and knowledge of the market.

Customers that I am aware have already stopped using Glyphosate since the IARC ruling:

Irvine Unified School District and several bay area cities and school districts.

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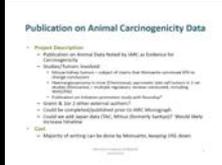
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Plaintiff Exhibit
0391



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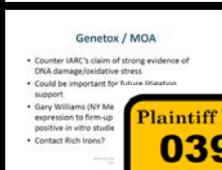
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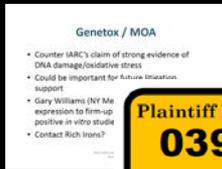
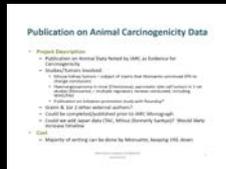
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2015

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Plaintiff Exhibit
0391



Mr. Johnson's Cancer



Jan.
2016

Mr. Johnson successfully gets permission to stop spraying Roundup at Benicia.

Mr. Johnson's Cancer

Jan.
2016

Mr. Johnson successfully gets permission to stop spraying Roundup at Benicia.

Aug.
2016

Mr. Johnson's cancer gets worse and worse.

Mr. Johnson's Cancer

Aug. 2016



Mr. Johnson's Cancer

Dec. 2016



Mr. Johnson's Cancer

Jan. 2017



Mr. Johnson's Cancer

June 2017



Mr. Johnson's Cancer

Nov. 2017



Mr. Johnson's Cancer

Jan. 2018



Mr. Johnson's Cancer

Absent a miracle, Mr.
Johnson will not live
to see 2020.

Mr. Johnson's Cancer



Dr. Nahban

Differential:

- ~~Age~~
- Race
- ~~Immunosuppressive drugs~~
- ~~Autoimmune disease~~
- Roundup exposures
- ~~Other chemical exposures~~
- ~~UV / Sun exposure~~
- ~~Viruses (HIV, HHV8, etc.)~~

Compensatory Damages

- Economic damages: \$2,253,209.32
- Non-economic damages
 - physical pain
 - mental suffering
 - loss of enjoyment of life
 - disfigurement
 - physical impairment
 - grief
 - anxiety
 - humiliation
 - emotional distress



Compensatory Damages

- Economic damages: \$2,253,209.32
- Non-economic damages

Past:

4 years: \$4,000,000

Future:

2-33 years: \$33,000,000

\$37,000,000

Compensatory Damages

- Economic damages: \$2,253,209.32
- Non-economic damages: \$37,000,000

TOTAL

\$39,253,209.23

3 Core Questions

YES

Can Roundup be a substantial contributing factor in causing cancer?

YES

Was Roundup a substantial contributing factor in causing Mr. Johnson's cancer?

3. Did Monsanto act with knowing disregard of human health?

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Jury Instruction

“Malice” means that Monsanto acted with intent to cause injury or that Monsanto’s conduct was despicable and was done with a willful and knowing disregard of the rights or safety of another. **A person acts with knowing disregard when he or she is aware of the probable dangerous consequences of his or her conduct and deliberately fails to avoid those consequences.**

3. Did Monsanto act with knowing disregard of human health?

Jury Instruction

Certain facts must be proved by clear and convincing evidence, which is a higher burden of proof. This means the party must persuade you that it is highly probable that the fact is true.

3. Did Monsanto act with knowing disregard of human health?

1. Dr. Parry's reports
2. Ghostwriting
3. Freedom to operate
4. Attacking IARC
5. Refusing to test
6. Targeting schools after IARC
7. MSDS v. consumer label
8. Never calling Mr. Johnson back

Jury Instruction

In view of Monsanto's financial condition, what amount is necessary to punish it and discourage future wrongful conduct?



Net Worth: **\$6.6 Billion**

Cash on hand: **\$3.1 Billion**

Interest (2%): **\$62 Million**

\$373,000,000

Verdict Form